

Effects of post-exercise sodium bicarbonate ingestion on acid-base balance recovery and time-to-exhaustion running performance: a randomised crossover trial in recreational athletes

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1 **ABSTRACT**

2 This study investigated the effect of post-exercise sodium bicarbonate (NaHCO₃) ingestion on acid-
3 base balance recovery and time-to-exhaustion (TTE) running performance. Eleven male runners
4 (stature, 1.80 ± 0.05 m; body mass, 74.4 ± 6.5 kg; maximal oxygen consumption, 51.7 ± 5.4 ml.kg⁻¹.min⁻¹)
5 participated in this randomised, single-blind, counterbalanced and crossover design study.
6 Maximal running velocity (v-VO_{2max}) was identified from a graded exercise test. During experimental
7 trials, participants repeated 100% v-VO_{2max} TTE protocols (TTE1, TTE2) separated by 40 min
8 following the ingestion of either 0.3 g.kg⁻¹ BM NaHCO₃ (SB) or 0.03 g.kg⁻¹ BM sodium chloride (PLA)
9 at the start of TTE1 recovery. Acid-base balance (blood pH and bicarbonate, HCO₃⁻) data were studied
10 at baseline, post-TTE1, after 35 min recovery and post-TTE2. Blood pH and [HCO₃⁻] were unchanged
11 at 35 min recovery (*p* > 0.05), but [HCO₃⁻] was elevated post-TTE2 for SB vs. PLA (+2.6 mmol.l⁻¹; *p*
12 = 0.005; *g* = 0.99). No significant differences were observed for TTE2 performance (*p* > 0.05), although
13 a moderate effect size was present for SB vs. PLA (+14.3 s; *g* = 0.56). Post-exercise NaHCO₃ ingestion
14 is not an effective strategy for accelerating the restoration of acid-base balance or improving subsequent
15 TTE performance when limited recovery is available.

16

17 **Key Words:** buffering; alkalosis; glycolytic activation; ergogenic aid; exercise capacity; nutrition.

18

19 **BULLET POINTS**

- 20 • Post-exercise sodium bicarbonate ingestion did not accelerate the restoration of blood pH or
21 bicarbonate after 35 minutes
- 22 • Performance enhancing effects of sodium bicarbonate ingestion may display a high degree of
23 inter-individual variation
- 24 • Small-to-moderate changes in performance were likely due to greater up-regulation of
25 glycolytic activation during exercise

26

27

28 INTRODUCTION

29 Repeated high-intensity training sessions and events require athletes to be able to maximally exert
30 themselves with often only limited recovery between exercise bouts (Barnett, 2006). The considerable
31 reliance on the ATP-PCr and glycolytic energetic systems results in the rapid accumulation of
32 biochemical metabolites such as inorganic phosphate, hydrogen cations (H^+) and lactate within
33 contracting muscle (Allen et al., 2008). Naturally occurring extracellular buffering mechanisms (e.g.,
34 blood bicarbonate, HCO_3^-) act to remove these H^+ from the muscle cytosol (Sahlin, 2014), however the
35 production of H^+ overwhelms the rate of neutralisation reactions, contributing towards reduced
36 intramuscular pH (Allen et al., 2008). These metabolically derived disruptions to acid-base balance (i.e.,
37 equilibrium between acidity and alkalinity of bodily fluids) have been linked to greater peripheral
38 fatigue via the inhibition of glycolytic flux (Messonnier et al., 2007), disruption of calcium ion cross-
39 bridge formation (Fitts, 2016) and reduced muscle excitability (Sostaric et al., 2006). Extracellular
40 buffering aids such as sodium bicarbonate ($NaHCO_3$) that protect against disturbances to acid-base
41 balance homeostasis could therefore prove beneficial to athletes performing repeated bouts of maximal
42 effort that are likely to exacerbate metabolic perturbation.

43 Pre-exercise ingestion of $0.3\text{ g}\cdot\text{kg}^{-1}$ BM $NaHCO_3$ enhances extracellular buffering capacity by
44 eliciting a metabolic alkalosis (i.e., increased blood $[HCO_3^-]$ of $\sim 5\text{-}7\text{ mmol}\cdot\text{l}^{-1}$) that allows for greater
45 H^+ efflux from the muscle (Messonnier et al., 2007), which may result in moderate ergogenic benefits
46 during high-intensity exercise (see review, Hadzic et al., 2019). This nutritional strategy also accelerates
47 the restoration of acid-base balance homeostasis within ~ 40 mins (Pruscino et al., 2008; Gough et al.,
48 2019a), but it is unclear whether the heightened HCO_3^- buffering response extends to subsequent athletic
49 performance (i.e., for athletes competing in multiple exercise events). Authors investigating the effect
50 of pre-exercise $NaHCO_3$ ingestion on repeated bout performance reported minimal ergogenic benefits
51 during wingate protocols (3x30 s; 15 min recovery) and swimming time-trials (2x200 m; 30 min
52 recovery) (Zabala et al., 2011; Pruscino et al., 2008). It is expected that pre-exercise ingestion resulted
53 in the enhanced HCO_3^- buffering capacity being completely utilised within the preliminary bout (i.e.,
54 from protecting against severe acid-base balance disturbances), which would explain blunted
55 performance benefits during subsequent exercise bouts.

56 One relatively novel strategy that accounts for this concern is post-exercise NaHCO₃ ingestion,
57 ensuring that a greater alkalotic response is present during recovery and subsequent exercise bouts.
58 Previous studies have demonstrated that NaHCO₃ ingested either immediately post-exercise or 30 min
59 after a fatiguing bout elevates blood pH (~0.10 units) and [HCO₃⁻] (~7-8 mmol.l⁻¹) following a 75-90
60 min recovery period (Gough et al., 2017; Gough et al., 2019), which subsequently improved time-to-
61 exhaustion (TTE) running (+70 s; *p* = 0.084) and cycling (+35 s; *p* = 0.007). Since the authors opted
62 for relatively long recovery windows, it was noted that blood pH and [HCO₃⁻] had not only recovered
63 prior to subsequent exercise, but instead reflected a state of pre-exercise metabolic alkalosis following
64 NaHCO₃ ingestion (~7.48 units, ~32 mmol.l⁻¹). Consequently, the ergogenic responses reported may
65 have represented the well-established pre-exercise application of NaHCO₃ ingestion. It is therefore
66 considered more appropriate to investigate the potential ergogenic effect of post-exercise NaHCO₃
67 when ingested during a shorter recovery period between two exercise bouts.

68 Performance enhancing effects following NaHCO₃ are not exclusively attributed to substantial
69 changes in blood pH and [HCO₃⁻] response (Siegler et al., 2016). This alkalotic state may also increase
70 glycolytic activation by preventing the allosteric inhibition of enzymes such as glycogen phosphorylase
71 and phosphofructokinase (Messonnier et al., 2007). Pre-exercise NaHCO₃ ingestion has been
72 demonstrated to augment glycolytic flux during high-intensity exercise protocols (da Silva et al., 2019;
73 Gurton et al., 2020). The effect of a post-exercise NaHCO₃ supplementation strategy on energy system
74 contribution during subsequent exercise remains underexplored, although authors have suggested
75 enhanced glycolytic flux via elevated post-exercise blood lactate (BLa⁻) concentration after a 75-90 min
76 recovery (Gough et al., 2017b; Gough et al., 2019b). It is conceivable that some athletes will have less
77 time to recover between high-intensity exercise bouts, therefore research should examine whether
78 NaHCO₃ ingested at the start of a shorter recovery period is still able to alter energy demand and
79 contribution from the glycolytic and oxidative systems.

80 From an applied perspective, athletes might be particularly attracted to nutritional strategies
81 that maximise ergogenic benefits during a second competition bout (i.e., track running/cycling, heat
82 then final; regional tennis, first and second round match). In these instances, ingestion of NaHCO₃
83 during a recovery period may prove more beneficial to subsequent performance. The purpose of this

84 study was to investigate the effect of post-exercise NaHCO₃ ingestion on the recovery of acid-base
85 balance (blood pH, [HCO₃⁻], base excess) within 35 mins and subsequent TTE running performance.

86

87 MATERIALS AND METHODS

88 Experimental approach to the problem

89 A block randomised, across subjects single-blind, placebo-controlled, counterbalanced, crossover
90 experimental design was implemented for this study. Experimental trial order was allocated by the
91 principal investigator using an online randomised number generator (www.randomization.com) and
92 blinded from other members of the research team. Participants attended three separate laboratory visits
93 in a 3-h post-prandial state to perform a graded exercise test and two experimental trials (separated by
94 5-7 d). All testing was completed at Surrey Human Performance Institute at the same time of day to
95 minimise the confounding effects of circadian rhythms on exercise performance (Reilly, 1990). The
96 final speed completed during the graded exercise test was classified as maximal velocity (100% v-
97 VO_{2max}) and used to prescribe TTE speed. During experimental trials, participants repeated 100% v-
98 VO_{2max} TTE protocols separated by 40 mins following the ingestion of either NaHCO₃ (SB) or sodium
99 chloride (NaCl; PLA) at the start of recovery. Vigorous exercise and the consumption of alcohol were
100 prohibited for 24-h prior to visits. Participants were instructed to record nutritional intake for 72-h prior
101 to attending the first laboratory visit using food diaries, which were checked by the principal
102 investigator and returned to participants to replicate before each experimental trial. Online databases
103 were used by the research team to determine the composition and nutritional content of foods.

104

105 Participants

106 A priori power calculation ($\beta = 0.8$; $\alpha = 0.05$) using previous partial eta-squared effect size data (η_p^2 ;
107 0.6) for post-exercise blood pH recovery (Gough et al., 2017a) revealed that a sample size of 10 would
108 allow detection of significant changes in blood pH (G*Power 3, Heinrich Heine, Düsseldorf, Germany).
109 Eleven recreationally trained runners (stature: 1.80 ± 0.05 m; body mass: 74.4 ± 6.5 kg; age: 31.0 ± 9.7
110 years; maximal oxygen consumption: 51.7 ± 5.4 ml.kg⁻¹.min⁻¹) were enrolled onto this study from a
111 local running club according to the CONSORT 2010 guidelines (Figure 1). All participants completed

112 at least 60 min vigorous exercise per week, had no history of hypertension (>140/90 mmHg) or renal
113 insufficiencies and had not ingested buffering agents within the previous 6 months. All study procedures
114 were reviewed and approved prior to commencing research by the **Faculty of Health and Medical**
115 **Sciences ethics committee at the University of Surrey (1366-FHMS-18)**. Participants gave informed
116 consent prior to participating in the study. Procedures were conducted in full accordance with the World
117 Medical Association's Declaration of Helsinki.

118

119 [INSERT **Figure 1** NEAR HERE]

120

121 **Procedures**

122 On the initial visit, baseline physiological measures were recorded (stature, body mass, blood pressure),
123 before participants performed a graded exercise test on a treadmill to determine 100% v-VO_{2max} for
124 TTE protocols. Gaseous exchange was collected using a breath-by-breath metabolic cart (Geratherm
125 Respiratory, Love Medical, Germany). After a 5 min warm-up at 6 km.h⁻¹ and 1% gradient, the speed
126 increased by 1 km.h⁻¹ every 1 min until volitional exhaustion was achieved (Price and Halabi, 2005).
127 This was deemed as the failure to maintain treadmill speed despite strong verbal encouragement. Heart
128 rate and rating of perceived exertion (RPE) were recorded at the end of each stage and volitional
129 exhaustion. Data were averaged over the peak 30 s for calculating VO_{2max} (Beltz et al., 2016) and 100%
130 v-VO_{2max} was classified as the final completed speed during the graded exercise test.

131 On arrival to the laboratory during experimental trial visits, participants completed visual
132 analog scales for gastrointestinal (GI) discomfort (0 mm = “no symptom at all”; 100 mm = “severest
133 symptom imaginable”) to quantify perceived severity of flatulence, abdominal discomfort (AD), gut
134 fullness (GF), bowel urgency rating (BUR) and diarrhoea (Gough et al., 2017a; Gurton et al., 2020).
135 Resting capillary blood samples (95 µl) were collected into end-to-end sodium-heparised capillary tubes
136 (EKF Diagnostics GmbH, Germany) and analysed for acid-base balance (blood pH, [HCO₃⁻], base
137 excess) using a handheld analyser (i-STAT, Abbott Point of Care, New Jersey). The i-STAT analyser
138 demonstrates excellent accuracy and precision for determination of blood pH and [HCO₃⁻] after
139 maximal exercise (intraclass correlation coefficients, ICC; $r = 0.88, 0.86$) (Dascombe et al. 2007).

140 Additional 20 µl blood samples were analysed for [BLa⁻] using an automatic analyser (Biosen C-Line,
141 EKF Diagnostic GmbH, Germany) that displays excellent precision (coefficient of variation, CV:
142 <1.5%; EKF Diagnostics, 2017). Participants perceived readiness to exercise (PRE) was measured,
143 before completing the first running TTE protocol (TTE1) at 100% v-VO_{2max} on the treadmill (1%
144 gradient). Breath-by-breath expired gaseous exchange and RPE were recorded throughout TTE
145 protocols. Capillary blood samples and visual analog scales were performed immediately post-TTE1,
146 before participants consumed either 0.3 g.kg⁻¹ BM NaHCO₃ (SB) or 0.03 g.kg⁻¹ BM NaCl (PLA) after
147 5 min recovery. These doses were chosen to replicate experimental design from previous studies
148 examining the effect of either pre- or post-exercise NaHCO₃ ingestion on recovery (Pruscino et al.,
149 2008; Gough et al., 2017b; Gough et al., 2019b). Experimental beverages were administered single-
150 blind as a chilled solution of 400 ml water and 100 ml orange squash to ensure palatability and taste-
151 matching (Lavender and Bird, 1989). Pilot testing revealed it was not possible to taste-match an
152 equimolar 0.21 g.kg⁻¹ BM NaCl dose (i.e., equal Na⁺ content between beverages). Participants
153 consumed beverages as fast as possible but were allocated a 5 min period if required (i.e., by 10 min
154 post-TTE1), before completing a supplement belief questionnaire to assess the single-blind design and
155 whether bias regarding the effect of NaHCO₃ was transferred onto participants (Gough et al. 2017a).
156 Capillary blood samples and visual analogue scales were repeated following 35 min recovery (30 min
157 post-ingestion). Participants were asked for their PRE and performed the second TTE protocol (TTE2)
158 at 100% v-VO_{2max} on the treadmill (40 min after TTE1). Additional blood samples were collected post-
159 TTE2, while visual analogue scales were completed post-exercise and for a 24-h post-laboratory phase.
160 Composite scores for GI discomfort were calculated at each time point from the sum of symptom
161 severity (i.e., scored out of 500 mm). An overview of experimental procedure is displayed in **Figure 2**.

162

163 [INSERT **Figure 2** NEAR HERE]

164

165 **Estimated energy system contribution calculations**

166 Absolute energy demand and contribution from the oxidative and glycolytic energetic systems were
167 estimated during TTE running protocols via non-invasive techniques (Milioni et al., 2017; Brisola et

168 al., 2015). Oxidative phosphorylation (W_{AER}) was calculated by subtracting resting oxygen
169 consumption (i.e., the average VO_2 value during the final 30 s of baseline) from the area under the
170 oxygen consumption curve for TTE protocols (di Prampero and Ferretti, 1999). Area under the curve
171 was calculated using the trapezoidal method. This approach provides reliable and valid estimations for
172 W_{AER} during high-intensity exercise (da Silva et al., 2019; Milioni et al., 2017). Glycolytic ($W_{[LA]}$) was
173 calculated from delta blood lactate (ΔBLa^-) during TTE protocols, according to the assumption that a
174 difference of 1 mmol.l^{-1} of BLa^- corresponded to 3 ml.kg^{-1} BM of O_2 (Milioni et al., 2017; Brisola et
175 al., 2015; Gurton et al., 2020), by using the following equation: $W_{[LA]} = \Delta BLa^- \times 3 \times \text{body mass (kg)}$.
176 The caloric quotient of 20.92 kJ was used to convert between absolute energy demand (in L of O_2) and
177 energy contribution (in kJ) for both energetic systems.

178

179 **Statistical analysis**

180 Normality and sphericity were assessed using Shapiro-Wilk and Mauchly tests, respectively. Any
181 violations were corrected via Greenhouse Geisser adjustments. Reproducibility of pre-trial nutritional
182 intake, TTE1 performance, perceived difficulty to exercise and initial acid-base balance perturbation
183 were determined from ICC's and categorised as either poor ($r \leq 0.40$), fair ($r = 0.40 - 0.59$), good ($r =$
184 $0.60 - 0.74$) or excellent ($r \geq 0.74$) (Atkinson and Nevill, 1998). Acid-base balance (primary outcome
185 measure) data were analysed using two-way (treatment x time) repeated measures ANOVA's, whereby
186 interaction effects were presented as a p value and effect size (η_p^2) (Olejnik and Algina, 2003).
187 Significant interactions were explored further by performing one-way repeated measures ANOVA
188 across treatments at *a priori* defined time points (i.e., post-TTE2) using the bonferroni correction factor.
189 The assumption of normal distribution was violated for GI discomfort therefore Friedman's two-way
190 ANOVA's were conducted as non-parametric alternative. Post-hoc Wilcoxon matched pair signed rank
191 tests were performed when significance was observed (median and Z score reported). Fisher's exact
192 test was used to assess the efficacy of the single-blind design. Paired t -tests were conducted on TTE2
193 performance (secondary outcome measure) and perceived difficulty to exercise. The within-subject CV
194 for the TTE1 protocol was calculated as 4.9% and used to interpret individual responses for changes in

195 TTE2 performance. Between treatment effect sizes were calculated by dividing mean differences by
196 pooled SD, before applying Hedges g bias correction (Lakens, 2013). These were interpreted as trivial
197 (< 0.20), small ($0.20 - 0.49$), moderate ($0.50 - 0.79$), or large (≥ 0.80) (Cohen, 1988). Data are presented
198 as Mean \pm SD (unless otherwise stated) and 95% confidence intervals (CI) reported for between
199 treatment statistical differences. Statistical significance was set at $p < 0.05$ and data were analysed using
200 SPSS v26 (SPSS Inc., IBM, USA).

201

202 RESULTS

203 Pre-trial nutritional intake

204 Participants demonstrated an excellent degree of repeatability for pre-trial nutritional intake during SB
205 and PLA trials for daily calories ($r = 0.89$, $p = 0.001$), carbohydrate ($r = 0.87$, $p = 0.002$), fat ($r = 0.83$,
206 $p = 0.005$) and protein ($r = 0.83$, $p = 0.014$).

207

208 Acid-base balance status

209 Initial acid-base balance perturbation demonstrated an excellent level of reproducibility during SB and
210 PLA trials for post-TTE1 blood pH ($r = 0.94$; $p < 0.001$), $[\text{HCO}_3^-]$ ($r = 0.95$; $p < 0.001$), base excess (r
211 $= 0.92$; $p < 0.001$) and $[\text{BLa}^-]$ ($r = 0.93$; $p < 0.001$). Significant two-way interactions (treatment \times time)
212 were observed for $[\text{HCO}_3^-]$ ($p = 0.002$; $\eta_p^2 = 0.413$) and $[\text{BLa}^-]$ ($p = 0.023$; $\eta_p^2 = 0.269$), but not for blood
213 pH ($p = 0.070$; $\eta_p^2 = 0.227$) or base excess ($p = 0.242$; $\eta_p^2 = 0.141$). Acid-base balance status was
214 unchanged at 35 min recovery ($p > 0.05$). Exploratory analysis on a small sub-set ($n = 6$) of blood pH
215 and $[\text{HCO}_3^-]$ data at 40 min recovery (i.e., immediately pre-TTE2; not an α priori defined time point)
216 revealed a larger magnitude of effect for SB vs. PLA (+0.03 units; +3.9 mmol.l^{-1}). Post-TTE2 $[\text{HCO}_3^-]$
217 was significantly elevated for SB vs. PLA ($p = 0.005$) and displayed a large effect size (+2.6 mmol.l^{-1} ;
218 CI: 1.0, 4.2; $g = 0.99$). Peak $[\text{BLa}^-]$ following TTE2 was significantly higher for SB vs. PLA ($p = 0.015$)
219 and displayed a large effect size (+1.4 mmol.l^{-1} ; CI: 0.3, 2.5; $g = 0.82$). Mean \pm SD values for changes
220 in acid-base balance status from baseline to post-TTE2 are displayed in **Figure 3 (A-D)**.

221

222 [INSERT **Figure 3 (A-D)** NEAR HERE]

223

224 **Time-to-exhaustion running performance**

225 Performance during TTE1 demonstrated an excellent degree of repeatability for SB and PLA trials ($r =$
226 0.95 ; $p < 0.005$; within-subject $CV = 4.9\%$). No significant differences were observed for TTE2
227 performance ($+14.3$ s; $p = 0.071$), although a moderate effect size was present for SB vs. PLA ($+9.7\%$;
228 $g = 0.56$). There was a large degree of individual variability for TTE2 performance, whereby mean
229 differences for SB vs. PLA ranged from -16.8 s (-14.2%) to $+61.2$ s ($+37.1\%$). In total, six out of eleven
230 participants reported improvements in TTE2 performance above the 4.9% test-retest variability. Sub-
231 analysis for these six ‘responders’ revealed a significant increase of $+31.2$ s for TTE2 performance
232 following SB compared to PLA ($+21.1\%$; $p = 0.007$; $g = 1.65$). Mean values and individual differences
233 for TTE2 performance are displayed in **Figure 4**.

234

235 [INSERT **Figure 4** NEAR HERE]

236

237 **Estimated energy system contribution**

238 Mean \pm SD values for energy demand (in L of O₂) and contribution (kJ) from oxidative and glycolytic
239 energetic systems are presented in **Table 1**. Excellent reproducibility was observed between SB and
240 PLA trials for energy contribution from W_{AER} ($r = 0.96$; $p < 0.001$) and $W_{[LA]}$ ($r = 0.92$; $p < 0.001$)
241 during TTE1. Energy contribution from $W_{[LA]}$ was significantly increased during TTE2 for SB vs. PLA
242 ($p = 0.032$) and displayed a moderate effect size ($+6.84$ kJ; CI: 0.7, 13.0; $g = 0.69$). In total, ten out of
243 eleven participants reported greater $W_{[LA]}$ contribution following SB vs. PLA. No significant differences
244 were observed for energy contribution from W_{AER} during TTE2 ($p > 0.05$), although a small effect size
245 was present for SB vs. PLA ($+11.20$ kJ; $g = 0.48$).

246

247 [INSERT **Table 1** NEAR HERE]

248

249 **Perceived difficulty of exercise**

250 Participants achieved a similar level of volitional exhaustion during TTE1 for SB and PLA trials as
251 good reproducibility was reported for peak RPE ($r = 0.66$; $p = 0.049$). No differences were observed
252 between SB and PLA for peak RPE during TTE2 ($p > 0.05$). No significant differences were reported
253 across treatments for PRE prior to TTE2 ($p > 0.05$), although a small effect size was present for SB vs.
254 PLA (+0.36 units; $g = 0.26$).

255

256 **Perceptual responses to experimental beverages**

257 Four participants identified SB, six were unsure on treatments, whilst one incorrectly identified SB as
258 PLA. Treatments were therefore considered successfully single-blinded and taste-matched (Fisher's
259 exact test, $p = 1.000$) and only a single participant directly suggested a poor taste.

260 In total, ten out of eleven participants experienced some GI discomfort for the SB trial (timing
261 and severity of peak symptom for each participant displayed in **Table 2**). **Composite scores for GI**
262 **discomfort revealed no symptoms at baseline, but symptom severity was mildly exacerbated for SB vs.**
263 **PLA at 35 min recovery (Mdn: 11 mm vs. 10 mm) and post-TTE2 (Mdn: 11 mm vs. 9 mm).** No
264 significant treatment effects were present at any time point for AD or GF ($p > 0.05$). No significant
265 treatment effects were observed at either 35 min recovery or post-TTE2 for flatulence, BUR and
266 diarrhoea (all $p > 0.05$). Ratings of GI discomfort were significantly exacerbated for SB vs. PLA during
267 the 24-h post-laboratory phase for flatulence (Mdn: 10 mm vs. 0 mm; $Z = 2.366$; $p = 0.018$), BUR (20
268 mm vs. 0 mm; $Z = 2.547$; $p = 0.011$) and diarrhoea (17 mm vs. 0 mm; $Z = 2.197$; $p = 0.028$). Out of the
269 five participants that failed to improve TTE2 performance for SB vs. PLA, only one directly reported
270 an ergolytic effect of GI discomfort.

271

272 [INSERT **Table 2** NEAR HERE]

273

274 **DISCUSSION**

275 This was the first study to investigate the effect of post-exercise NaHCO_3 ingestion on blood pH and
276 $[\text{HCO}_3^-]$ recovery after 35 min and subsequent TTE running performance. No changes in acid-base
277 balance status were reported for SB vs. PLA after the 35 min recovery period, however both $[\text{HCO}_3^-]$

278 and $[BLa^-]$ were elevated post-TTE2. Post-exercise $NaHCO_3$ ingestion did not significantly improve
279 TTE2 performance, although a moderate effect size and a high degree of inter-individual variation was
280 observed. Glycolytic energy system contribution was moderately increased during TTE2 for SB vs.
281 PLA. Lastly, post-exercise $NaHCO_3$ ingestion resulted in moderate-to-severe GI symptoms after TTE2
282 and for the 24-h post-laboratory phase, therefore athletes should remain cautious when using this
283 strategy to improve performance.

284 The novel finding from this study was that $NaHCO_3$ ingested after a fatiguing exercise bout
285 failed to accelerate the recovery of blood pH, $[HCO_3^-]$ and base excess to 'neutral' levels within 35 min
286 when compared to PLA. These results contrast findings from pre-exercise ingestion studies suggesting
287 that $NaHCO_3$ accelerated the restoration of acid-base balance homeostasis within 30-40 min (Pruscino
288 et al., 2008; Gough et al., 2019a). This discrepancy is most likely explained by the pre-exercise ingestion
289 strategy previously employed, since this would have resulted in the enhanced buffering response being
290 utilised during initial exercise (i.e., protecting against severe acid-base balance disturbances), in turn
291 allowing for blood pH and $[HCO_3^-]$ to recover to baseline status 'faster'. Indeed, Gough et al. (2017b)
292 reported that $NaHCO_3$ administered following TTE cycling improved acid-base balance within 90 min
293 compared to a placebo (blood pH, ~ 0.10 units; $[HCO_3^-]$, ~ 7.0 mmol.l⁻¹). Nonetheless, as the authors
294 waited until ~ 30 min post-exercise to administer $NaHCO_3$, it was observed that blood pH and $[HCO_3^-]$
295 had already naturally returned close to baseline (~ 7.42 ; ~ 25.0 mmol.l⁻¹), therefore their results may
296 represent the widely regarded alkalotic effect of pre-exercise supplementation.

297 Our findings complement those from Gough et al. (2019b) demonstrating that post-exercise
298 $NaHCO_3$ ingestion from a state of metabolic acidosis resulted in a greater $[HCO_3^-]$ after 75 min recovery
299 ($+8.6$ mmol.l⁻¹). The relatively unchanged acid-base balance status at 35 min recovery herein can be
300 attributed to a blunted extracellular buffering response since orally administered HCO_3^- had likely not
301 been fully absorbed within the duodenum (Gough et al., 2017a). It is expected that substantial changes
302 in enhanced HCO_3^- buffering only commenced during the second TTE protocol, which is demonstrated
303 by the elevated $[HCO_3^-]$ for post-TTE2 following SB, thus offering one possible explanation for the
304 moderate effect size reported for TTE2 performance. These findings suggest however that the ingestion

305 of NaHCO₃ immediately post-exercise is not an effective strategy for accelerating the restoration of
306 acid-base balance homeostasis when only a limited recovery period is available.

307 The performance enhancing effects of NaHCO₃ ingestion on TTE running were considerably
308 blunted when compared to previous post-exercise NaHCO₃ studies (Gough et al., 2017b; Gough et al.,
309 2019b), instead more closely representing the equivocal benefits reported during repeated bout exercise
310 protocols (Zabala et al., 2011; Pruscino et al., 2008). The discrepancy between results is attributed to a
311 dampened extracellular buffering response, whereby all participants ended the recovery period in a
312 relatively 'neutral' acid-base balance status (i.e., opposed to a metabolic alkalosis from previous post-
313 exercise studies). Despite the continual rise in [HCO₃⁻] during the warm-up prior to TTE2 that was
314 demonstrated by our sub-set of data for 40 min recovery, it is unlikely that differences in [HCO₃⁻] for
315 SB vs. PLA consistently achieved the 5.0-6.0 mmol.l⁻¹ minimal buffering threshold that has been
316 theorised to maximise potential performance benefits (Carr et al., 2011). These changes in TTE2
317 performance did however display a large degree of inter-individual variation, whereby a significant
318 'true' effect of NaHCO₃ was observed for the six participants (i.e., 'responders') that improved TTE2
319 duration above the 4.9% within-subject CV for the TTE protocol. Considering that large variability in
320 performance responses following NaHCO₃ ingestion is common (Saunders et al., 2014), likely due to
321 between participant differences in HCO₃⁻ absorption kinetics (Gough et al. 2017a), it is recommended
322 that athletes first trial the supplement within training regimes, before then applying to a competitive
323 'real-world' situation if positive results are obtained. Nonetheless, as three of these 'responders'
324 identified SB (from a breaking of study blinding due to GI symptoms) and reported higher PRE (~2
325 units) prior to TTE2, it is also possible that improvements were partially accounted to an 'expectancy
326 of ergogenicity' following NaHCO₃ ingestion (Higgins and Shaabir, 2016). Additional work should
327 now investigate the inter-individual variation of performance enhancing effects following post-exercise
328 NaHCO₃ ingestion on more robust time-trial protocols that are less likely to be influenced by
329 psychological bias.

330 Post-exercise NaHCO₃ ingestion elevated W_[LA] energy contribution during subsequent
331 exercise, which agrees with previous pre-exercise ingestion findings (Gurton et al., 2020; da Silva et
332 al., 2019) and offers additional mechanistical insight to explain changes in TTE performance. Despite

333 the limited recovery period, an acute NaHCO₃ dose induced a sufficient change in alkalotic state to
334 augment total glycolytic flux during TTE2, which was likely caused by up-regulated lactate-H⁺ co-
335 transport from intramuscular environments via the monocarboxylate transporter (Messonnier et al.,
336 2007). The elevated post-TTE2 [BLa⁻] indirectly demonstrates increased activation of the glycolytic
337 energetic system following NaHCO₃ ingestion, although the magnitude of increase was lower than from
338 previous studies (Gough et al., 2017b; Gough et al., 2019b). One explanation for this discrepancy relates
339 to the diminished extracellular buffering response as [HCO₃⁻] was only ~3-4 mmol.l⁻¹ higher at the start
340 of TTE2 for SB vs. PLA, whereas Gough et al. (2017b) induced a substantial metabolic alkalosis prior
341 to the second exercise bout ([HCO₃⁻]: ~7 mmol.l⁻¹ higher), resulting in a greater number of H⁺ that could
342 be neutralised. Based on the 1:1 stoichiometry of the HCO₃⁻ and H⁺ reaction, and assuming that total
343 blood volume was ~5 L, then SB from this current study would have only neutralised an additional ~15
344 mmol.l⁻¹ of H⁺ compared to PLA (opposed to ~30 mmol.l⁻¹ from Gough et al., 2017b), therefore
345 providing justification for our dampened up-regulation of glycolytic contribution. It is important to note
346 though, our results may partially reflect greater lactate efflux from muscles, instead of an increased
347 glycolytic flux (da Silva et al., 2019). Considering that participants also reported **small, unexpected**
348 **increases for W_{AER} during TTE2, which were partly attributed to the greater total work performed for**
349 **SB vs. PLA (i.e., increased running distance)**, it is possible that we slightly overestimated the ‘true’
350 effect of NaHCO₃ on energy system contribution. Future research should investigate whether NaHCO₃
351 ingested within recovery directly up-regulates glycolytic flux (i.e., from invasive muscle biopsies
352 techniques) during subsequent controlled work-rate/duration exercise protocols.

353 Moderate-to-severe GI symptoms following the dissociation of NaHCO₃ within gastric acid
354 have been widely documented (Cameron et al., 2010; Carr et al., 2011). Our results demonstrated **mildly**
355 **exacerbated GI discomfort for SB at 35 min recovery compared to baseline**, which is in agreement to
356 one post-exercise ingestion study (Gough et al., 2017b), but not the other (Gough et al., 2019b). **These**
357 **disparities might be attributed to differences in assessment protocols (i.e., visual analogue scales vs. 11-**
358 **point likert scales)**, or the established inter-individual variation for GI discomfort, whereby some
359 athletes report greater gastric tolerability to NaHCO₃ ingestion than others (Heibel et al., 2018). GI
360 discomfort was moderately exacerbated for SB vs. PLA post-TTE2 (i.e., reflecting worse symptoms

361 during exercise), however only a single participant suggested these side-effects impacted TTE2
362 performance, therefore our results reaffirm that GI discomfort might not directly exhibit an ergolytic
363 effect (Higgins et al., 2013; Gough et al., 2018). One novel finding was that post-exercise NaHCO₃
364 ingestion resulted in severe GI discomfort for the 24-h post-laboratory phase (i.e., $n = 5$, diarrhoea), in
365 theory proving detrimental to athletes during future training and competition. Athletes suffering from
366 severe GI discomfort could opt for delivery strategies that bypass gastric side-effects, such as
367 administration via enteric-coated capsules which adequately reduce severity and occurrence of GI
368 symptoms (Hilton et al., 2020a), whilst still eliciting a sufficient, **albeit delayed metabolic alkalosis**
369 **(>6.0 mmol.l⁻¹; peak response, 90-110 min)** that improves exercise performance (Hilton et al., 2020b).
370 **Additional work is required to elucidate the efficacy of enteric-coated NaHCO₃ capsules when only a**
371 **limited amount of time is available between exercise bouts.**

372 There are some methodological limitations in the present study. Firstly, a single-blind study
373 design is considered sub-optimal for nutritional intervention research, however this was purely due to
374 logistical reasons (i.e., lack of research staff). Additional control measures were put in place such as the
375 standardisation of verbal encouragement during TTE protocols (e.g., “keep going” every 15 s) and the
376 use of a supplement belief questionnaire (Gough et al., 2017a) that for the most part indicated we
377 blinded NaHCO₃ from participants. Another limitation was our inability to directly quantify glycolytic
378 flux and contribution from the ATP-PCr energetic system. Due to the importance of measuring blood
379 pH and [HCO₃⁻] immediately post-exercise, we were unable to identify a clear excess-post oxygen
380 consumption curve after TTE exercise, therefore ATP-PCr calculations were removed from analysis.
381 Lastly, the 35 min recovery was potentially too short for post-exercise NaHCO₃ ingestion to have a
382 substantial effect on acid-base balance recovery, although previously published data (Gough et al.,
383 2017a) suggests that 0.3 g.kg⁻¹ BM NaHCO₃ increases [HCO₃⁻] by ~5.0 mmol.l⁻¹ within 40 min. This
384 length of recovery was chosen as it best reflects the time required for blood pH and [HCO₃⁻] to naturally
385 return close to baseline (Gough et al., 2017b; Pruscino et al., 2008), which allowed us to investigate the
386 effect of post-exercise NaHCO₃ ingestion on the restoration of acid-base balance homeostasis and
387 whether this improved subsequent high-intensity performance.

388

389 **CONCLUSION**

390 The ingestion of NaHCO₃ immediately after an initial fatiguing exercise bout is not an effective strategy
391 for accelerating the restoration of acid-base balance homeostasis (blood pH, HCO₃⁻, base excess) when
392 only a limited recovery period is present. This was attributed to a blunted extracellular buffering
393 response, most likely as orally administered HCO₃⁻ had not been fully absorbed within the duodenum.
394 Post-exercise NaHCO₃ ingestion did not significantly improve TTE2 performance, although the
395 moderate effect size and high degree of variability advocate that coaches and practitioners administer
396 NaHCO₃ on an individual basis. Small-to-moderate changes in TTE2 performance were likely due to
397 enhanced HCO₃⁻ buffering and the up-regulation of glycolytic activation specifically during TTE2
398 exercise. NaHCO₃ ingestion resulted in severe post-exercise GI discomfort that may prove detrimental
399 to athletes during subsequent training bouts or competition events. Future research should investigate
400 the application of alternative NaHCO₃ administration strategies (e.g., enteric-coated capsules) on the
401 recovery of acid-base balance, GI discomfort and repeated bout exercise performance.

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404

405 **Competing interests statement**

406 The authors declare that there are no competing interests related to the nutritional supplement or
407 outcomes of the study.

408

409 **Contributors' statement**

410 WG and DK designed the study. WG, HM and DK collected the data. WG performed data analysis and
411 interpretations. WG prepared the majority of manuscript, HM, LG and DK also contributed. All authors
412 reviewed the manuscript and provided feedback.

413

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416

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Figure captions

Figure 1 CONSORT 2010 reporting guidelines for research flowchart.

Figure 2 Overview of study design and experimental procedures. Abbreviations: *v-VO_{2max}*, maximal running velocity; *SB*, sodium bicarbonate; *PLA*, placebo; *GI*, gastrointestinal; *BLa⁻*, blood lactate; *BG*, blood gas; *PRE*, perceived readiness to exercise; *TTE1/2*, time-to-exhaustion protocol one/two.

Figure 3 (A-D) Mean \pm SD changes in acid-base balance status (**A**, blood bicarbonate; **B**, blood pH; **C**, base excess; **D**, blood lactate). Some error bars are removed for clarity. * denotes significantly higher for SB vs. PLA ($p < 0.05$). Dashed horizontal lines represent baseline/normal acid-base balance status.

Figure 4 Mean differences and inter-individual variation (dashed lines between data points) for subsequent time-to-exhausting running protocols.