

# **Vascular, inflammatory, and perceptual responses to hot water immersion; impacts of water depth and temperature in young healthy adults**

## **Original article**

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**Author contributions:**

Campbell Menzies, Tom Cullen, Neil Clarke, Christopher Pugh, and Doug Thake were responsible for the conception and design of the study. Campbell Menzies, Charles Steward, and Tom Cullen were responsible for data acquisition, while all authors assisted in interpretation of the data. All authors contributed to drafting or revision of the written work, approved the final version and agree to be accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. All persons designated as authors qualify for authorship, and all those who qualify for authorship are listed.

**What is the central question of this study?**

How do the acute vascular, inflammatory, and perceptual responses differ with changes in water temperature, immersion depth, and rectal temperature using three different 30-minute hot water immersion protocols?

**What is the main finding and its importance?**

Increases in water temperature and immersion depth during 30-minute immersion protocols do not result in different effects on inflammation but have distinct region-specific arterial haemodynamic effects. Beneficial physiological responses may be accompanied with less favourable perceptual responses demonstrating a potential trade-off that should be considered during protocol implementation.

**Abstract**

Repeated hot water immersion can improve cardiovascular health; however, the respective effects of distinct immersion protocols remain unclear. Twenty-two healthy adults completed three 30-minute hot water immersion bouts of different water temperatures and immersion depths (40-Shoulder, 42-Waist, 40-Waist) in a randomised cross-over design. Vascular, inflammatory, and perceptual responses were collected via brachial and superficial femoral artery ultrasound, venous blood sampling, and perceptual scales. Rectal temperature increased less in the 40-Waist ( $\Delta 0.5 \pm 0.1$  °C) condition than the other conditions (40-Shoulder:  $\Delta 0.9 \pm 0.3$  °C, 42-Waist:  $\Delta 0.9 \pm 0.3$  °C,  $p < 0.001$ ). Arm skin temperature increased more in the 40-Shoulder ( $\Delta 5.2 \pm 1.9$  °C) condition than the other conditions (40-Waist:  $\Delta 2.6 \pm 1.0$  °C, 42-Waist:  $\Delta 3.6 \pm 1.1$  °C,  $p < 0.001$ ), whilst thigh temperature had a greater increase in the 42-Waist ( $8.6 \pm 1.3$  °C) condition than either the 40-Waist ( $7.8 \pm 0.2$  °C) or 40-Shoulder ( $\Delta 7.8 \pm 1.0$  °C) conditions ( $p < 0.001$ ). Brachial artery shear rate was greatest post-immersion following the 40-Shoulder condition (40-Shoulder:  $\Delta 121 \pm 94$  1/s, 42-Waist:  $\Delta 47 \pm 73$  1/s, 40-Waist:  $\Delta -21 \pm 41$  1/s,  $p < 0.001$ ) whereas superficial femoral artery shear rate was largest following the 42-Waist condition (40-Shoulder:  $\Delta 143 \pm 61$  1/s, 42-Waist:  $196 \pm 85$  1/s, 40-Waist:  $131 \pm 93$  1/s,  $p < 0.001$ ). IL-6 ( $p = 0.16$ ) and cortisol ( $p = 0.83$ ) responses did not differ between conditions. Perceptual responses were more favourable in the 40-Waist condition. Taken together, these data demonstrate the distinct region-specific arterial responses align with increases in local skin temperature to alterations in hot water immersion protocols, whilst showing that beneficial physiological responses may be accompanied with less favourable perceptual responses.

## Introduction

Passive heating, shares many physiological responses with exercise (Cullen *et al.*, 2020) and repeated exposure can result in enhanced cardiorespiratory fitness (Bailey *et al.*, 2016; Hesketh *et al.*, 2019), vascular adaptation (Carter *et al.*, 2014b; Bailey *et al.*, 2016; Brunt *et al.*, 2016), and a range of adaptive responses in skeletal muscle (Kim *et al.*, 2020); including angiogenesis (Hesketh *et al.*, 2019), mitochondrial biogenesis (Hafen *et al.*, 2018), and improved glucose metabolism (Ely *et al.*, 2019; Hoekstra *et al.*, 2018). These adaptive processes are underpinned, in part, by the acute shear stress (Carter *et al.*, 2014b) and inflammatory (Hoekstra *et al.*, 2018) responses to heating, as well as activation of intracellular signalling pathways, such as increased AMPK activity (Liu & Brooks, 2011). Whilst a considerable evidence-base exists to support the prescription of specific and optimal exercise protocols (American College of Sports Medicine, 2020), in comparison, passive heating research is in its infancy, with little consensus on best practice for heating protocol optimisation in relation to mode, duration, temperature, or body coverage. Indeed, duration, temperature, and body coverage combine to determine the overall heating dose making it difficult to isolate their individual effects when interpreting the physiological effects of a heating protocol. Accordingly, the acute physiological responses, such as potential differences in thermo-physiological, haemodynamic, or inflammatory effects between heating protocols are yet to be defined and require systematic investigation to provide valuable insight when designing and implementing novel acute and longitudinal interventions.

Acute increases in blood flow and shear stress are key stimuli for both micro- and macro-vascular chronic adaptation following passive heating (Carter *et al.*, 2014b, 2014a). Indeed, acute changes in shear rate and blood pressure after a single exercise or hot water immersion exposure positively correlated with subsequent adaptation to repeated usage (Dawson *et al.*, 2018; Roxburgh *et al.*, 2023). Local limb temperature is a key driver for increased peripheral blood flow to that limb (Chiesa *et al.*, 2016; Koch Esteves *et al.*, 2024; Watanabe *et al.*, 2024), however whole body heating can increase limb temperature and blood flow in unheated limbs (Heinonen *et al.*, 2011; Chaseling *et al.*, 2023). Indeed, lower body heating can evoke systemic effects, as evidenced by increased brachial artery shear rate and flow-mediated dilation (Amin *et al.*, 2021; Cheng *et al.*, 2021). These systemic effects may be triggered by whole body thermoregulatory responses that occur following increases in core body temperature (Low *et al.*, 2011). Accordingly, although acute arterial haemodynamic responses appear to be evoked by increases in both local and core body temperature, the magnitude of response varies across vascular beds (Cheng *et al.*, 2021; Francisco *et al.*, 2021; Hoekstra *et al.*, 2021) and is likely

significantly influenced by the body coverage of the heating stimulus. Moreover, blood flow continues to increase when rectal temperature is clamped during prolonged hot water immersion (Francisco *et al.*, 2021). This means that studies investigating the effects of rectal temperature on haemodynamic responses during passive heating are likely confounded by duration (e.g. Chiesa *et al.*, 2016). Manipulation of heating dose through different water temperatures and immersion depths may allow for duration and rectal temperature-matched protocols to be assessed to elucidate systemic and localised haemodynamic responses.

Similar to arterial haemodynamic responses, many inflammatory and endocrine responses appear sensitive to increases in core body temperature (Rhind *et al.*, 2004). Acute elevations in inflammatory cytokines, such as interleukin-6 (IL-6), are suggested to be important in the subsequent anti-inflammatory response and protecting against chronic inflammatory and metabolic diseases (Nash *et al.*, 2023). Although mixed findings of acute IL-6 responses to passive heating have been observed, with some (Hoekstra *et al.*, 2020, 2021; Mansfield *et al.*, 2021) but not all (Monroe *et al.*, 2021; Gibson *et al.*, 2025) studies showing an acute elevated IL-6 concentration, it has recently been suggested that IL-6 may increase following passive heating in a dose-dependent manner (Hoekstra *et al.*, 2020). However, the specific factors that influence ‘dose’ are currently ill-defined. Heat-induced IL-6 release from skeletal muscle has been demonstrated *ex vivo* (Welc *et al.*, 2012; Obi *et al.*, 2017), suggesting that increases in muscle rather than core temperature may be the primary stimulus for the acute inflammatory response to passive heating. Indeed, human studies have shown that, IL-6 responses to local and whole-body heating appear similar despite smaller increases in rectal temperature with local heating (Hoekstra *et al.*, 2021). Therefore, the potential inflammatory responses to passive heating may differ with heating duration, body coverage, or temperature but this requires further investigation using carefully controlled experimental protocols, with systematic manipulation of whole body and local temperature, to enable the relevant stimuli to be isolated.

Whilst passive heating may offer physiological benefits, some protocols can be uncomfortable, difficult to tolerate, and result in negative effects, such as orthostatic hypotension (Steward *et al.*, 2023). Mitigation strategies may be effective in reducing these effects (Steward *et al.*, 2023); however, they may also reduce the desired physiological responses. Indeed, upper body cooling with lower body heating improves perceptual responses compared to whole body heating, but at the cost of attenuated increases in inflammatory and haemodynamic responses (Hoekstra *et al.*, 2021). Accordingly, the balance between the desired physiological responses

and the tolerability or potential negative effects is an important consideration when prescribing any passive heating intervention.

The present study aimed to systematically manipulate water temperature and immersion depth to investigate limb specific vascular, acute inflammatory, and perceptual responses to hot water immersion protocols of matched-duration and eliciting similar increases in rectal temperature. It was hypothesised that: (i) All immersion protocols would increase arterial shear rate in both submerged and non-submerged limbs, with higher water temperatures augmenting these responses, and the greatest increase being observed in the local arteries of submerged limbs;; (ii) The IL-6 response would be greater with larger increases in rectal temperature; and (iii) Thermal comfort, desired exit time, and dizziness upon standing would differ between conditions.

## Methods

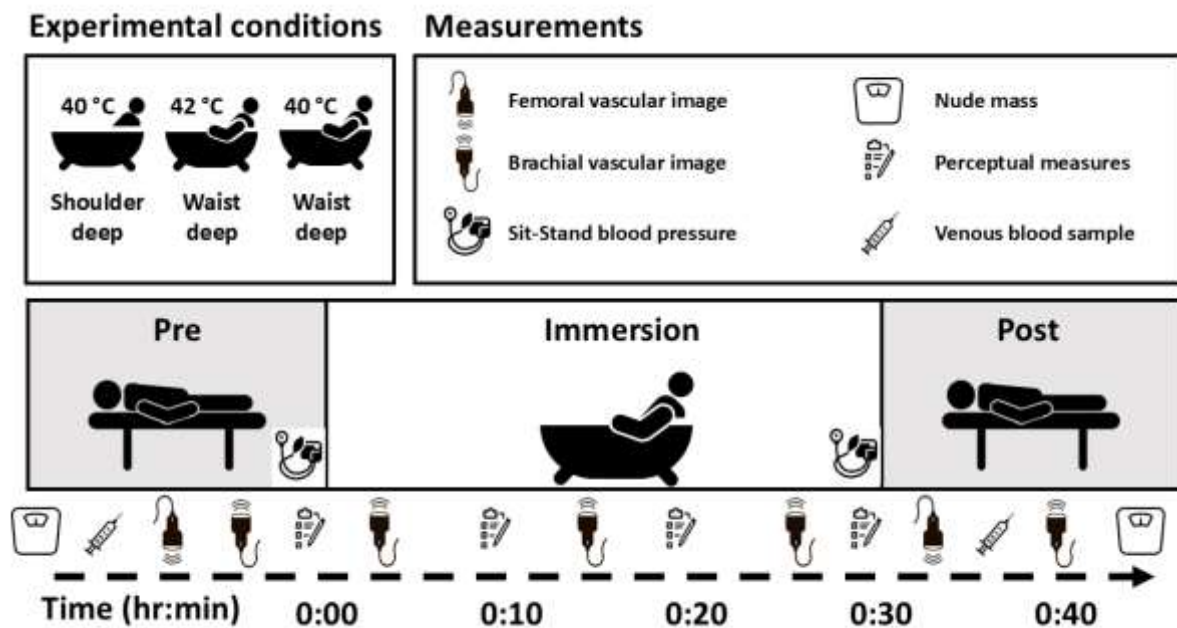
*Participants* – Based on previously observed effects for acute vascular responses ( $d = 0.74$ ; Chaseling et al., 2023) and IL-6 ( $d = 0.65$ ; Mansfield et al., 2021) between different heating protocols, the present study was powered *a priori* to detect an effect size of  $d = 0.65$ , with an alpha of 0.05 and an 80% power resulting in a required sample size of 22. Accordingly, 22 healthy adults (13 males:  $29 \pm 6$  years,  $80.3 \pm 14.4$  kg,  $1.78 \pm 0.09$  m,  $25.3 \pm 4.4$  kg/m<sup>2</sup>. 9 females:  $27 \pm 6$  years,  $62.1 \pm 8.8$  kg,  $1.64 \pm 0.05$  m,  $22.9 \pm 2.2$  kg/m<sup>2</sup>) were briefed and provided written consent to participate in the present study. Ethical approval was provided by the Coventry University Ethics committee (P146084) and conformed to the *Declaration of Helsinki*, except for prior registration in a database. Participants were aged 18-40 years old, non-smokers, with no history of cancer or metabolic illness, and had not been exposed to regular passive heating for at least three months prior to participating in the study.

*Experimental design* – Participants attended the laboratory ( $20.0 \pm 1.4$  °C,  $53 \pm 10\%$  relative humidity) on three occasions to complete 30 minutes of hot water immersion. The three experimental conditions consisted of (i) 40 °C shoulder-deep immersion (40-Shoulder) ( $40.0 \pm 0.1$  °C), (ii) 42 °C waist-deep immersion (42-Waist) ( $42.0 \pm 0.1$  °C), or (iii) 40 °C waist-deep immersion (40-Waist) ( $40.0 \pm 0.1$  °C) and were completed in a randomised cross-over design. These conditions were chosen such that the 40-Shoulder and 42-Waist conditions elicited similar increases in rectal temperature based on previous research (Hoekstra *et al.*, 2018; Mansfield *et al.*, 2021) and pilot testing within our laboratory. A 30-minute duration was chosen as the shortest duration previously shown to elicit adaptation with repeated hot water

immersion exposures (Bailey *et al.*, 2016). For the waist-deep conditions, participants sat on an 18 cm stool without immersion of their arms in a water depth of ~36 cm. In the shoulder-deep condition participants sat on the bottom of the hot tub with their arms submerged in a water depth of 47 – 52 cm, adjusted according to the participants height, and moved to the stool for ~3 minutes with non-submersion of their arms after 5, 15, and 25 minutes for the assessment of blood pressure and vascular imaging. Water temperature was constantly monitored using a thermistor (Grant instruments, UK) and maintained using a hot tub heating generator (LazySpa Majorca, Bestway International Ltd, Hong Kong) set to 40 °C, with addition and removal of hot or cold water as required to maintain temperature and depth.

Visits were separated by a minimum of 48 hours and conducted at the same time of day ( $\pm 2$  h), with the order of experimental conditions determined by a randomisation sequence in Excel. To control for effects of the menstrual cycle on thermoregulatory and endothelial responses (Hashimoto *et al.*, 1995; Charkoudian *et al.*, 2017), sessions were completed between days 1 and 7 after their self-reported onset of menses for eumenorrheic participants ( $n = 6$ ), or during the 7 inactive days for participants taking oral contraceptive pills ( $n = 2$ ), with one participant having an intra-uterine device and no menstrual cycle.

*Experimental protocol* – Prior to arrival in the laboratory, participants were instructed to refrain from strenuous exercise for 24 hours, be fasted for > 6 hours and refrain from caffeine consumption for 12 hours. Upon arrival, participants commenced with the experimental protocol (Figure 1).



**Figure 1.** Schematic of experimental design representing the three experimental conditions, time course of immersion, and measurements taken.

*Thermo-physiological measures* – Rectal temperature was monitored at five-minute intervals throughout using a rectal thermometer self-inserted 10 cm beyond the anal sphincter (Grant instruments, UK). Heart rate was recorded using a heart rate chest strap (H10, Polar, Kempele, Finland). Nude mass (Seca; Bodycare, UK) was recorded before and after the experimental protocol to estimate whole body sweat rates. In a subset of six participants (5 male, 1 female), skin temperature was measured (DS1921H iButton, Maximum, Dallas Semiconductor Corp., USA) on the upper arm and thigh adjacent to the location of the probe location for vascular imaging.

*Perceptual measures* – Thermal sensation, and thermal comfort were measured on scales ranging from +5 (hot and very comfortable, respectively) to -5 (cold and very uncomfortable, respectively) modified from Epstein & Moran (2006), with thermal desirability measured from +3 (a significantly lower temperature would be better) to -3 (a significantly higher temperature would be better) modified from Oi et al. (2012). During immersion, participants were asked for ratings of thermal sensation for above and below the water separately, thermal comfort overall, and thermal desirability of the water. Symptoms of dizziness, confusion, nausea, tiredness, and headaches were monitored on a 0 – 10 scale, with the scale anchored at 0 - no symptoms, 3 – mild symptoms, 5 – moderate symptoms, 7 – severe symptoms, and 10 – have



to stop (Coris *et al.*, 2006). Participants were also asked every five minutes whether under conditions outside of the laboratory, they would choose to exit the water. Following completion of all the conditions, participants ranked the three conditions in order of favourite to least favourite.

*Orthostatic hypotension* – Orthostatic hypotension was measured as previously described (Steward *et al.*, 2023) and defined according to clinical guidelines (Freeman *et al.*, 2011) as a reduction of  $\geq 20$  mmHg in systolic blood pressure (SBP) or  $\geq 10$  mmHg in diastolic blood pressure (DBP) upon standing when compared with values obtained at baseline, before the immersion period. Briefly, blood pressure (M3 Blood pressure monitor, Omron, Kyoto, Japan) was recorded in duplicate whilst seated on a chair at baseline following a 5-minute period of quiet rest. Upon standing, participants reported symptoms of dizziness on the same 0 – 10 scale used for symptoms of heat illness (Coris *et al.*, 2006) whilst a single blood pressure measurement was taken. Participants were instructed to stand up gradually and give a rolling dizziness score (i.e. repeated verbalisation of perceived dizziness every few seconds) with maximum dizziness recorded. Similarly, maximum heart rate upon standing was recorded due to the role of heart rate maintaining cardiac output to counteract reductions in blood pressure during orthostasis (Convertino, 2014).

*Vascular imaging* – The superficial femoral artery was imaged pre and three minutes post immersion. Brachial artery imaging was performed pre and nine minutes post immersion. Resting images were performed following at least 10 minutes of supine rest. During immersion, brachial artery diameter, blood flow, and shear rate were measured after 5, 15, and 25 minutes, whilst participants rested their arm on the edge of the hot tub. Vascular imaging was performed using a 15-MHz multifrequency linear array probe attached to a high-resolution duplex ultrasound machine (T3300; Tersaon, Burlington, MA). Images were taken following optimisation of the longitudinal B-mode image of the lumen-arterial interface, with simultaneous Doppler velocity assessments collected using the smallest possible insonation angle (always  $< 60^\circ$ ). For each visit, once optimal image acquisition was achieved, the probe location was marked on the skin to ensure imaging of the same arterial section.

Analysis of arterial diameter, blood flow and shear rate was performed using custom-designed edge-detection and wall-tracking software, as previously described elsewhere (Woodman *et al.*, 2001). This semi-automated software is independent of investigator bias and has greater reproducibility than manual methods (Woodman *et al.*, 2001), and in the present study had a

between visit coefficient of variation (CV) at rest of  $3.9 \pm 3.3\%$  and  $4.7 \pm 3.5\%$  for brachial and superficial femoral artery diameter, respectively. Blood flow was calculated as lumen cross-sectional area x Doppler velocity and shear rate (as an estimate of shear stress without viscosity) as four times the mean blood velocity/vessel diameter.

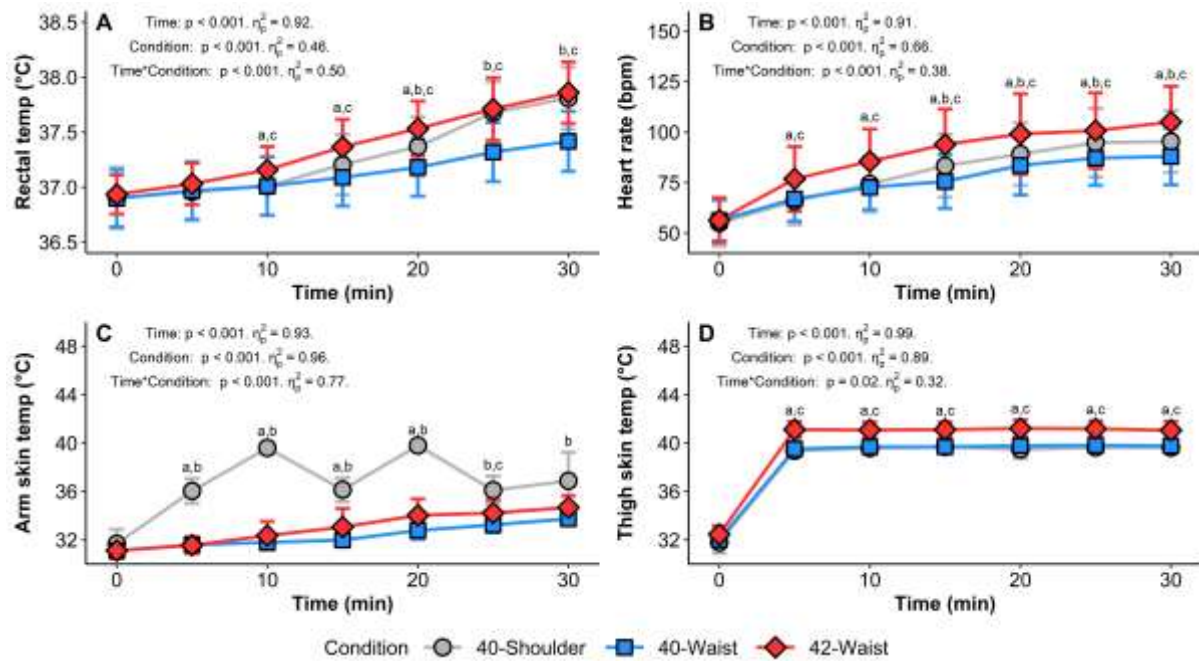
*Blood sampling and analysis* –Venous blood samples (~10 mL) were obtained pre and ~5 minutes post immersion through repeated venepuncture. Blood samples were split for the determination of haematocrit and haemoglobin (Hb 201+ System, Haemocue, Sweden), with the remaining blood spun at 3000 xg for 10 minutes in an *EDTA* vacutainer before aliquoting the plasma which was stored at -80 °C until later analysis. IL-6 and cortisol were analysed using commercially available ELISA kits (D6050 & KGE008B, respectively. R&D systems, Abingdon, United Kingdom). Samples were diluted (DY997, R&D Systems) in a ratio of 1:60 for the cortisol assay to ensure concentrations were within the dynamic range of the assay. Concentrations were determined using interpolation of a 4-parameter standard curve (GraphPad Prism, San Diego, Calif., USA) before being adjusted for changes in plasma volume calculated according to Dill & Costill (1974). To minimise variation between assays, all samples from an individual participant were analysed in the same assay and in the present study, the intra-assay CV for these assays were  $8.8 \pm 8.1\%$  and  $3.4 \pm 2.8\%$  for IL-6 and cortisol, respectively. Due to sampling issues, blood samples were not obtained for two participants (1 male, 1 female), resulting in a sample of n = 20 for these outcomes.

*Statistical analysis* –Data are reported as mean  $\pm$  standard deviation, or median (lower quartile, upper quartile) unless stated otherwise, with mean difference and 95% confidence intervals (95%CI) presented as mean difference [lower limit, upper limit] when describing differences between conditions. Analysis was conducted, unless otherwise stated, in Rstudio using the functions and packages stated below, with an example of the code found in the supplementary material. Statistical significance was accepted as  $\alpha < 0.05$ . Perceptual data (Thermal comfort, Thermal sensation, Thermal desirability, Condition preference) were examined within each condition for an effect of time using the *freidman.test* function from the *stats* package to conduct a Friedman test. Differences between conditions were also analysed using a Friedman test and where differences were detected, post-hoc testing was performed using the *pairwise.wilcox.test* function from the *stats* package, with Bonferroni adjustments for multiple comparisons. Count data (desired exit time, presence of orthostatic hypotension) were analysed using the *chisq.test* and *chisq.post.hoc* functions from the *stats* and *fifer* packages, respectively to perform Chi-squared tests with Fishers exact tests applied where significant effects of

condition were found. Change in nude mass and plasma volume were analysed using a one-way repeated measures ANOVA whilst, all other data were examined using a two-way (Time\*Condition) repeated measures ANOVA using the *anova\_test* function from the *Rstatix* package, with effect sizes for main effects presented as partial eta squared ( $\eta_p^2$ ). Violations of normality were not assessed due to the risks associated with non-Gaussian models and the robustness of ANOVA in violations of this assumption (Knief & Forstmeier, 2020). Where an interaction effect was identified, post-hoc analysis between conditions at the separate time points were performed using the *pairwise\_t\_test* function from the *Rstatix* package, with Bonferroni adjustments for multiple comparisons. Although not the focus of this study, sex differences were analysed by adding Sex as a between subjects variable into a three-way (Sex\*Time\*Condition) ANOVA, with the results available in the Supplementary material.

## Results

*Thermo-physiological measures* – Rectal temperature increased in all conditions but was lower at the end of immersion in the 40-Waist condition compared to both the 40-Shoulder (mean difference: 0.4 [0.3, 0.5] °C,  $p < 0.001$ ) and 42-Waist conditions (mean difference: 0.4 [0.4, 0.5] °C,  $p < 0.001$ ), which were similar (mean difference: 0.1 °C [-0.2, 0.1] °C,  $p = 1.0$ ), (Figure 2A). Similarly, changes in nude mass were not different between 40-Shoulder ( $\Delta -0.4 \pm 0.2$  kg) and 42-Waist ( $\Delta -0.4 \pm 0.2$  kg,  $p = 0.70$ ), but with a smaller reduction in 40-Waist ( $\Delta -0.3 \pm 0.2$  kg, mean difference vs 40-Shoulder: -0.5 [-0.6, -0.4] kg,  $p < 0.001$ , mean difference vs 42-Waist: -0.4 [-0.5, -0.4] kg,  $p = 0.003$ ) resulting in a significant main effect of condition ( $p < 0.001$ ,  $\eta_p^2 = 0.40$ ). In contrast, heart rate was higher at the end of immersion in the 42-Waist condition than the 40-Shoulder condition (mean difference: 10 [5, 14] bpm,  $p = 0.001$ ), which was in turn higher than the 40-Waist condition (mean difference: 7 bpm [3, 12] bpm,  $p = 0.003$ ) (Figure 2B). Arm skin temperature increased during immersion and was greater in the 40-Shoulder condition compared to the 40-Waist condition throughout immersion and compared to the 42-Waist for the first 20 minutes of immersion (Figure 2C). Arm skin temperature was also higher in the 42-Waist condition, compared to the 40-Waist condition after 25 minutes. Thigh skin temperature increased during immersion and was greater in the 42-Waist condition than the 40-Shoulder or 40-Waist conditions, which were similar (Figure 2D).



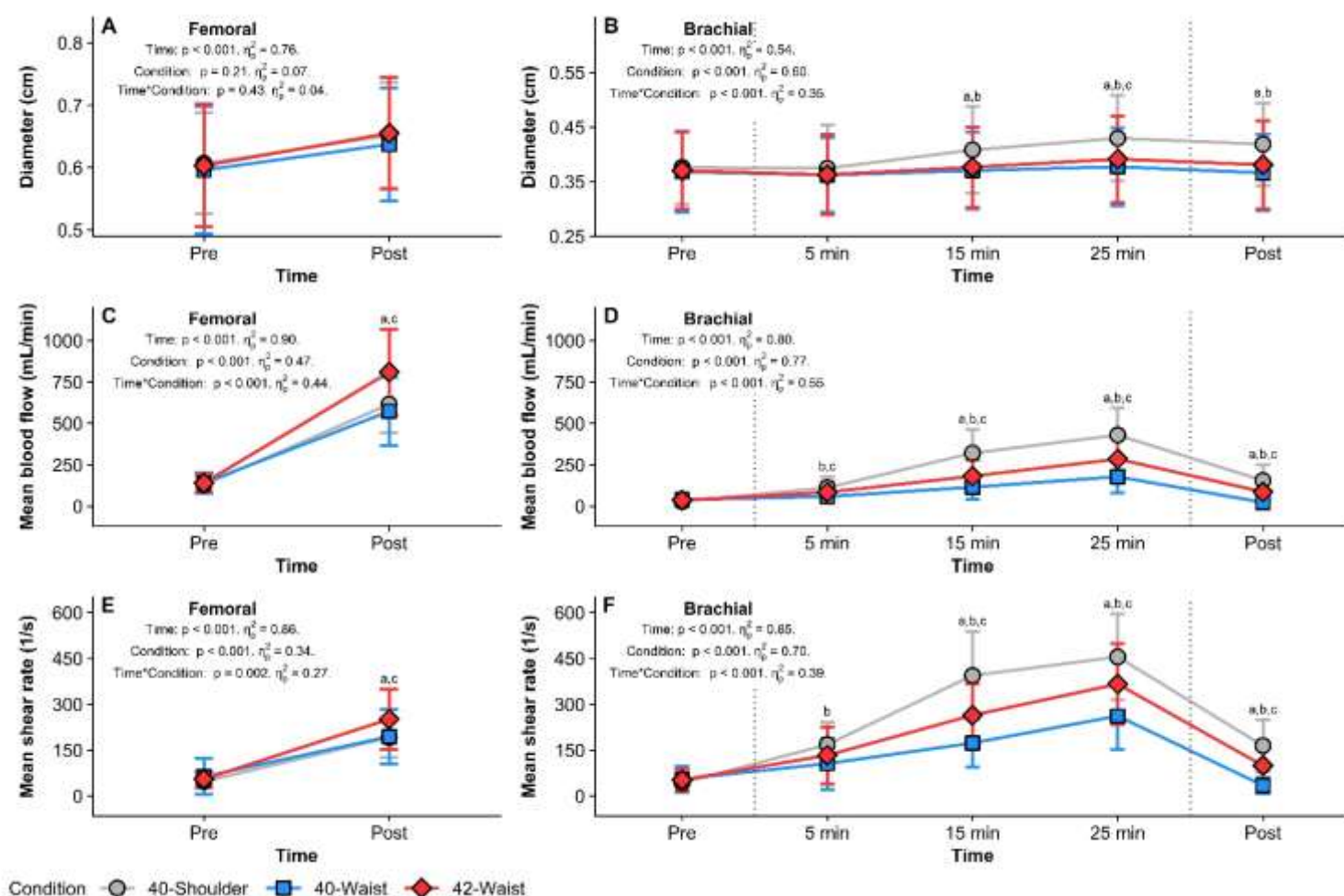
**Figure 2.** Mean thermo-physiological responses to each condition over time. A - Rectal temperature, B - Heart rate, C – Arm skin temperature, D – Thigh skin temperature. Significance is denoted as follows: <sup>a</sup>40-Shoulder vs 42-Waist, <sup>b</sup>40-Shoulder vs 40-Waist, and <sup>c</sup>42-Waist vs 40-Waist.

*Blood pressure responses* - Systolic and diastolic blood pressure reduced during immersion in all conditions, with the greatest reduction in systolic blood pressure, but smallest reduction in diastolic blood pressure, observed in the 40-Waist condition (Table 1). There was no difference in systolic blood pressure between conditions post-immersion; however, diastolic blood pressure was different between all conditions, being lowest in the 40-Shoulder condition (mean difference vs 40-Waist: -9 [-12, -6] mmHg,  $p < 0.001$ . mean difference vs 42-Waist: -4 [-7, -1] mmHg,  $p = 0.04$ ), and highest in the 40-Waist condition (mean difference vs 42-Waist: 5 [2, 8] mmHg,  $p = 0.02$ ).

*Superficial femoral artery responses* - There was a similar increase in diameter in all conditions (Figure 3A). However, there was a difference between conditions for mean blood flow (Figure 3C), with greater values observed post-immersion in the 42-Waist condition than either the 40-Shoulder (mean difference: 193 [109, 278] mL/min,  $p < 0.001$ ) or 40-Waist conditions (mean difference: 273 [154, 319] mL/min,  $p < 0.001$ ). These differences were also observed post-immersion for mean shear rate (Figure 3E; mean difference 42-Waist vs 40-Shoulder: 60 [30, 91] 1/s,  $p = 0.003$ . 42-Waist vs 40-Waist: 57 [35, 79] 1/s,  $p < 0.001$ ) and antegrade shear rate (Table 1; mean difference 42-Waist vs 40-Shoulder: 59 [29, 89] 1/s,  $p = 0.003$ . 42-Waist vs

40-Waist: 55 [33, 76] 1/s,  $p < 0.001$ ). There were no observed differences in these variables between the 40-Shoulder and 40-Waist conditions. In contrast, there was no difference between conditions in the significant reduction of retrograde shear rate observed following immersion (Table 1).

*Brachial artery responses* – Diameter increased during immersion peaking at 25 minutes before falling post-immersion (Figure 3B), with larger values observed at 25 minutes in the 40-Shoulder condition than either the 42-Waist (mean difference: 0.38 [0.27, 0.50] cm,  $p < 0.01$ ) or 40-Waist conditions (mean difference: 0.52 [0.40, 0.64] cm,  $p < 0.01$ ), which also differed (mean difference: 0.14 [0.04, 0.24] cm,  $p = 0.05$ ). Similar responses were observed, with largest values observed at 25 minutes, for mean blood flow (Figure 3D; mean differences: 40-Shoulder vs 42-Waist: 145 [97, 193] mL/min,  $p < 0.001$ . 40-Shoulder vs 40-Waist: 251 [199, 303] mL/min,  $p < 0.001$ . 42-Waist vs 40-Waist: 106 [72, 141] mL/min,  $p < 0.001$ ), mean shear rate (Figure 3F; mean differences: 40-Shoulder vs 42-Waist: 89 [34, 143] 1/s,  $p = 0.01$ . 40-Shoulder vs 40-Waist: 193 [134, 253],  $p < 0.001$ . 42-Waist vs 40-Waist: 105 [57, 153] 1/s,  $p = 0.001$ ), and antegrade shear rate (Figure 3D; mean differences: 40-Shoulder vs 42-Waist: 81 [32, 129] 1/s,  $p = 0.01$ . 40-Shoulder vs 40-Waist: 182 [125, 238] 1/s,  $p < 0.001$ . 42-Waist vs 40-Waist: 101 [53, 149] 1/s,  $p = 0.001$ ). Retrograde shear rate was different between all conditions post-immersion (Table 1) being negligible in the 40-Shoulder condition and highest in the 40-Waist condition (mean differences: 40-Shoulder vs 42-Waist: 8 [4, 11] 1/s,  $p < 0.001$ . 40-Shoulder vs 40-Waist: 16 [11, 22] 1/s,  $p < 0.001$ . 42-Waist vs 40-Waist: 9 [3, 14] 1/s,  $p = 0.02$ ).



**Figure 3.** Mean arterial diameter, mean blood flow and mean shear rate responses across time for each condition for the superficial femoral (A, C, E) and brachial (B, D, F) arteries, respectively. The data between the vertical dotted lines in B, D, & F were obtained from the brachial artery during immersion, whilst participants were sat upright, whilst Pre and Post measures across all panels were obtained with participants supine. Significance is denoted as follows: <sup>a</sup>40-Shoulder vs 42-Waist, <sup>b</sup>40-Shoulder vs 40-Waist, and <sup>c</sup>42-Waist vs 40-Waist.

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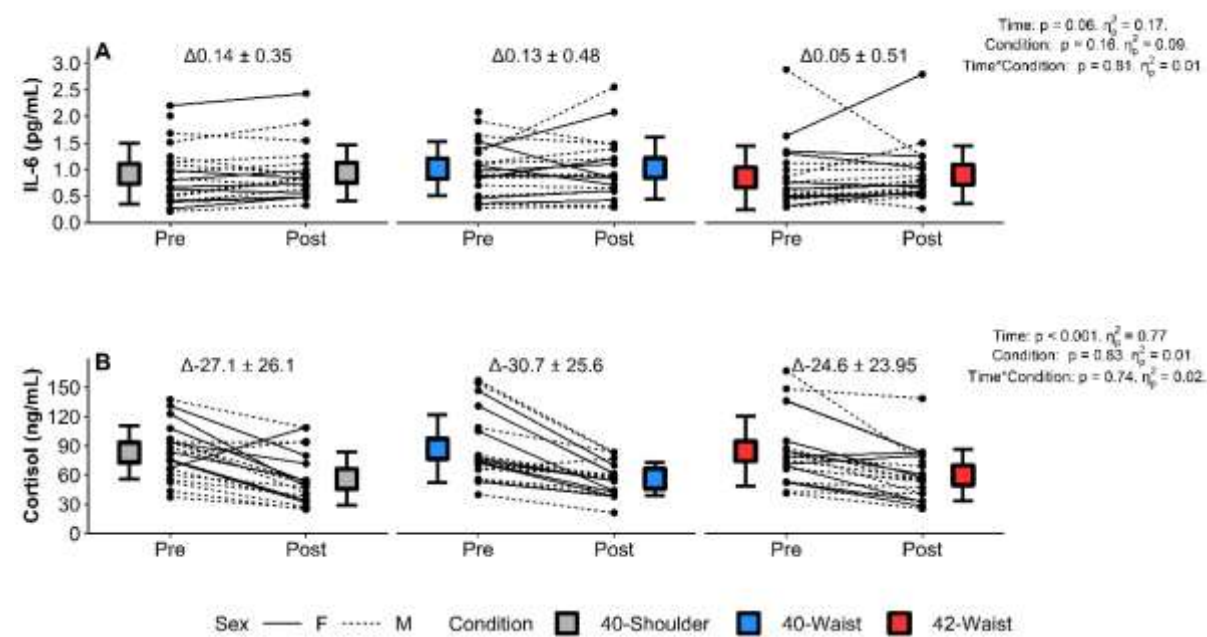
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**Table 1.** Summary of mean antegrade and retrograde responses in the brachial and superficial femoral arteries, and mean blood pressure for each condition over time. Significance is denoted as follows: <sup>a</sup>40-Shoulder vs 42-Waist, <sup>b</sup>40-Shoulder vs 40-Waist, and <sup>c</sup>42-Waist vs 40-Waist.

Variable	Condition	Pre	5 min	15 min	25 min	Post	Statistics
Systolic blood pressure (SBP) (mmHg)	40-Shoulder	118 ± 10	109 ± 13 <sup>b</sup>	112 ± 14	113 ± 12	123 ± 10	Time: $p < 0.001$ , $\eta_p^2 = 0.66$
	42-Waist	118 ± 8	107 ± 14	112 ± 14	114 ± 14 <sup>c</sup>	124 ± 8	Condition: $p = 0.60$ , $\eta_p^2 = 0.13$
	40-Waist	119 ± 11	105 ± 14 <sup>b</sup>	107 ± 17	108 ± 17 <sup>c</sup>	123 ± 11	Time*Condition: $p = 0.02$ , $\eta_p^2 = 0.10$
Diastolic blood pressure (DBP) (mmHg)	40-Shoulder	69 ± 6	61 ± 8	56 ± 10	54 ± 7 <sup>b</sup>	58 ± 9 <sup>a,b</sup>	Time: $p < 0.001$ , $\eta_p^2 = 0.68$
	42-Waist	70 ± 5	60 ± 9	58 ± 10	57 ± 8	62 ± 9 <sup>a,c</sup>	Condition: $p = 0.003$ , $\eta_p^2 = 0.25$
	40-Waist	70 ± 6	59 ± 7	61 ± 9	58 ± 9 <sup>b</sup>	67 ± 6 <sup>b,c</sup>	Time*Condition: $p < 0.001$ , $\eta_p^2 = 0.16$
Femoral antegrade shear rate (1/s)	40-Shoulder	77 ± 26	-	-	-	196 ± 60 <sup>a</sup>	Time: $p < 0.001$ , $\eta_p^2 = 0.83$
	42-Waist	84 ± 33	-	-	-	255 ± 95 <sup>a,c</sup>	Condition: $p < 0.001$ , $\eta_p^2 = 0.33$
	40-Waist	92 ± 56	-	-	-	200 ± 85 <sup>c</sup>	Time*Condition: $p < 0.001$ , $\eta_p^2 = 0.29$
Femoral retrograde shear rate (1/s)	40-Shoulder	-30 ± 12	-	-	-	-5 ± 7	Time: $p < 0.001$ , $\eta_p^2 = 0.81$
	42-Waist	-28 ± 12	-	-	-	-3 ± 7	Condition: $p = 0.60$ , $\eta_p^2 = 0.02$
	40-Waist	-28 ± 15	-	-	-	-6 ± 7	Time*Condition: $p = 0.48$ , $\eta_p^2 = 0.03$
Brachial antegrade shear rate (1/s)	40-Shoulder	59 ± 35	184 ± 69 <sup>b</sup>	395 ± 143 <sup>a,b</sup>	457 ± 139 <sup>a,b</sup>	165 ± 85 <sup>a,b</sup>	Time: $p < 0.001$ , $\eta_p^2 = 0.87$
	42-Waist	64 ± 38	167 ± 76	276 ± 94 <sup>a,c</sup>	376 ± 121 <sup>a,c</sup>	107 ± 66 <sup>a,c</sup>	Condition: $p < 0.001$ , $\eta_p^2 = 0.66$
	40-Waist	68 ± 41	140 ± 76 <sup>b</sup>	201 ± 73 <sup>b,c</sup>	275 ± 102 <sup>b,c</sup>	51 ± 29 <sup>b,c</sup>	Time*Condition: $p < 0.001$ , $\eta_p^2 = 0.38$
Brachial retrograde shear rate (1/s)	40-Shoulder	-16 ± 14	-16 ± 20 <sup>a,b</sup>	-1 ± 5 <sup>a,b</sup>	-2 ± 4 <sup>b</sup>	0 ± 1 <sup>a,b</sup>	Time: $p < 0.001$ , $\eta_p^2 = 0.36$
	42-Waist	-12 ± 10	-34 ± 32 <sup>a</sup>	-12 ± 13 <sup>a</sup>	-9 ± 20	-8 ± 9 <sup>a,c</sup>	Condition: $p < 0.001$ , $\eta_p^2 = 0.41$
	40-Waist	-13 ± 10	-34 ± 29 <sup>b</sup>	-28 ± 36 <sup>b</sup>	-13 ± 21 <sup>b</sup>	-17 ± 13 <sup>b,c</sup>	Time*Condition: $p < 0.001$ , $\eta_p^2 = 0.16$

NB: Participants were supine for Pre and Post measurements but sat upright during immersion at 5, 15, and 25 min.

*Venous blood measures* - Increases in plasma volume were observed following each bout of immersion and were similar between all conditions (40-Shoulder:  $\Delta 11 \pm 16\%$ . 42-Waist:  $\Delta 9 \pm 12\%$ . 40-Waist:  $\Delta 9 \pm 10\%$ .  $p = 0.80$ .  $\eta_p^2 = 0.01$ ). IL-6 did not differ over time, or between conditions (Figure 4A), whilst cortisol reduced post-immersion, but did not differ between conditions (Figure 4B).



**Figure 4.** Mean plasma IL-6 (A) and Cortisol (B) concentrations pre and post immersion for each condition. Connected lines and smaller dots represent individual responses for each participant. Abbreviations: IL-6 – Interleukin-6.

*Perceptual measures* – Summary data for all perceptual measures are shown in Table 2. Thermal sensation (above and below the water), thermal comfort, and thermal desirability were similar between 40-Shoulder and 42-Waist for 20 minutes but differed at 30 minutes where participants felt hotter, less comfortable, and desired a lower water temperature in the 42-Waist condition. In the 40-Waist condition, thermal sensation (above the water), thermal comfort, and thermal desirability were different from the other two conditions for the first 20 minutes of immersion with participants feeling cooler, more comfortable, and having less desire to reduce the water temperature. However, these variables were similar to the 40-Shoulder condition after 30 minutes, when participants in this condition had moved to the stool and removed their arms from the water. Symptoms of heat illness were not largely prevalent in any condition (Table 3). After 20 minutes of immersion, there was a difference between conditions for the number of participants who had indicated they would choose to exit the water outside



of laboratory conditions (40-Shoulder: n = 12. 42-Waist: n = 13, 40-Waist: n = 5. p = 0.03), however comparisons between conditions were not identified with post-hoc testing. A further three participants in each condition indicated they would exit the water prior to the 30-minute time point and did not alter the statistical comparison between conditions. There was no clear difference in participant preference for each protocol (p = 0.11). However, 40-Shoulder was rated as the favourite condition by 13 participants, compared to seven in the 40-Waist condition and two in the 42-Waist condition. Inversely, six participants rated 40-Shoulder as their least favourite condition, which was fewer than both the 42-Waist (n = 9), and 40-Waist (n = 7) conditions.

**Table 2.** Median perceptual responses over time for each condition. Significance is denoted as follows: \* significant effect of time within variable, <sup>a</sup>40-Shoulder vs 42-Waist, <sup>b</sup>40-Shoulder vs 40-Waist, and <sup>c</sup>42-Waist vs 40-Waist.

Variable	Condition	0	10	20	30
Thermal sensation (below water) (a.u)	40-Shoulder*	-1 (-2.75, -0.25)	3 (1.25, 3)	4 (3, 4) <sup>b</sup>	4 (2, 4)
	42-Waist*	-1 (-2, 0)	3 (2, 4) <sup>c</sup>	4 (3.25, 4) <sup>b,c</sup>	4 (4, 5) <sup>c</sup>
	40-Waist*	-1 (-2, -0.25)	2 (1, 3) <sup>c</sup>	3 (2, 3) <sup>c</sup>	3 (3, 4) <sup>c</sup>
Thermal sensation (above water) (a.u)	40-Shoulder*	-1 (-2.75, -0.25)	1 (0.25, 2) <sup>b</sup>	3 (1.25, 3) <sup>b</sup>	2 (1, 3) <sup>a</sup>
	42-Waist*	-1 (-2, 0)	1 (0, 2) <sup>c</sup>	3 (2, 3) <sup>c</sup>	4 (3, 4.75) <sup>a,c</sup>
	40-Waist*	-1 (-2, -0.25)	0 (-1, 0) <sup>b,c</sup>	0.5 (0, 1) <sup>b,c</sup>	1 (1, 2) <sup>c</sup>
Thermal comfort (a.u)	40-Shoulder*	0 (-1, 0.75)	2 (0, 3)	-1 (-2, 1) <sup>b</sup>	0 (-1.75, 1.75) <sup>a</sup>
	42-Waist*	0 (-1, 1.75)	0.5 (-1, 2)	-1 (0, -2) <sup>c</sup>	-2 (-4, -1) <sup>a,c</sup>
	40-Waist*	0 (-1, 0.75)	2 (1, 3)	2 (0, 2.75) <sup>b,c</sup>	0 (-1, 1) <sup>c</sup>
Thermal desirability (of the water) (a.u)	40-Shoulder*	-	0 (0, 0) <sup>b</sup>	1 (0, 1) <sup>b</sup>	1 (0, 1) <sup>a</sup>
	42-Waist*	-	0 (0, 1) <sup>c</sup>	1 (1, 2) <sup>c</sup>	2 (1, 2) <sup>a,c</sup>
	40-Waist*	-	0 (-0.75, 0) <sup>b,c</sup>	0 (0, 0) <sup>b,c</sup>	0.5 (0, 1) <sup>c</sup>

388

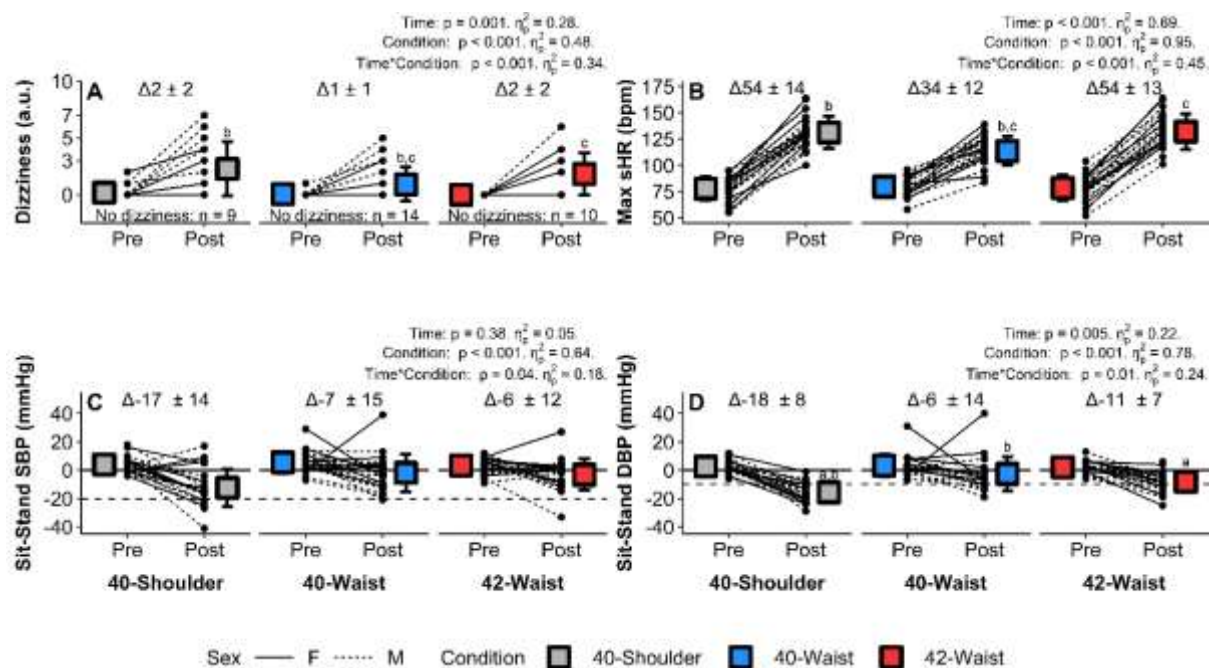
**Table 3.** Frequency of symptoms of heat illness reported in each condition.

Condition	Confusion	Dizziness	Headaches	Nausea	Tiredness	Total
40-Shoulder	22/0/0/0/0	21/0/0/1/0	22/0/0/0/0	22/0/0/0/0	21/0/1/0/0	108/0/1/1/0
42-Waist	22/0/0/0/0	20/1/1/0/0	20/2/0/0/0	22/0/0/0/0	20/1/1//0	104/2/4/0/0
40-Waist	22/0/0/0/0	22/0/0/0/0	22/0/0/0/0	22/0/0/0/0	20/1/1//0	108/1/1/0/0

Values are shown as the peak response across the protocol across all participants for scores of 0/1-2/3-4/5-6/7-10.

*Orthostatic hypotension* - The prevalence of orthostatic hypotension was significantly different between conditions (p = 0.001), with a higher prevalence in the 40-Shoulder condition (17/22; 77%) than the 40-Waist condition (5/22; 23%. p = 0.002), but no statistical difference with the 42-Waist condition (9/22; 41%. p = 0.09). These differences were largely driven by changes in Sit-Stand DBP (Figure 5D) rather than SBP (Figure 5C), with lower values observed post-

immersion in the 40-Shoulder condition than either the 42-Waist (mean difference: -7 [-3, -10] mmHg,  $p = 0.001$ ) or 40-Waist conditions (mean difference: -13 [-7, -19] mmHg,  $p = 0.004$ ). Lower values of dizziness upon standing were observed post-immersion in the 40-Waist condition than in the 40-Shoulder (mean difference: -1 [-1, -2] a.u.,  $p = 0.008$ ) and 42-Waist conditions (mean difference: -1 [0, -1] a.u.,  $p = 0.01$ ) (Figure 5A). Maximum heart rate upon standing displayed a similar pattern with an increase in heart rate following all immersion conditions, but lower values post-immersion in the 40-Waist condition than the 40-Shoulder (mean difference: -18 [-12, -23] bpm,  $p < 0.001$ ) or 42-Waist conditions (mean difference: -18 [-14, -23] bpm,  $p < 0.001$ ) (Figure 5B).



**Figure 5.** Mean responses upon standing pre and post immersion for each condition. A – Standing dizziness, B – Maximum heart rate upon standing, C – Sit-Stand SBP, D – Sit-Stand DBP. Significance is denoted as follows: <sup>a</sup>40-Shoulder vs 42-Waist, <sup>b</sup>40-Shoulder vs 40-Waist, and <sup>c</sup>42-Waist vs 40-Waist. Connected lines and smaller dots represent individual responses for each participant. Dashed horizontal lines represent the clinical threshold for orthostatic hypotension in C – SBP (-20 mmHg), and D - DBP (-10 mmHg). Abbreviations: max sHR - Maximum heart rate upon standing, DBP – Diastolic blood pressure, SBP – Systolic blood pressure.

## Discussion

The present study aimed to investigate acute arterial haemodynamic, inflammatory, and perceptual responses following three different 30-minute hot water immersion protocols, which manipulated water temperature, immersion depth, and rectal temperature to isolate their effects. The main findings were that: (i) Limb submersion results in the greatest increases in arterial shear (i.e. greater increases in the brachial artery with 40-Shoulder than 42-Waist, despite similar increases in rectal temperature). However, higher water temperatures augment the response in both submerged (i.e. femoral artery responses between 42-Waist and 40-Waist) and non-submerged limbs (i.e. brachial artery responses between 42-Waist and 40-Waist). Increased shear rate was accompanied by greater elevations in local skin temperature highlighting the importance of local temperature in arterial haemodynamic responses; (ii) 30-minutes of hot water immersion was insufficient to elicit significant changes in circulating IL-6 concentrations. In contrast, cortisol concentrations reduced following immersion but did not differ between conditions; (iii) Increasing the heating stimulus through either immersion depth, or water temperature increases symptoms of dizziness, and maximum heart rate, whilst reducing the decrease in diastolic blood pressure upon standing following immersion and decreasing thermal comfort during immersion.

### *Vascular responses*

The present study provides novel insight into the acute effects of hot water immersion, demonstrating that shear rate responses are most pronounced within submerged limbs, and that higher water temperatures augment this response. Higher thigh skin temperature corresponded to increased shear rate in the superficial femoral artery, while increased arm skin temperature was accompanied by elevated brachial artery shear rate. In contrast, despite similar increases in rectal temperature between the 40-Shoulder and 42-Waist conditions increases in shear rate were both protocol and limb specific, with increases in rectal temperature having no effect on superficial femoral artery shear rate when thigh skin temperatures were similar between the 40-Shoulder and 40-Waist conditions. These findings support the importance of local temperature on arterial haemodynamic responses to heating (Heinonen *et al.*, 2011; Chiesa *et al.*, 2016; Koch Esteves *et al.*, 2024; Watanabe *et al.*, 2024) and emphasise that elevations in core body temperature are not the primary driver of vascular responses. Previous data have demonstrated sequentially greater increases in rectal temperature (in response to a single prolonged heating bout) result in progressive increases in superficial femoral artery shear rate, with a  $\sim 30$  1/s increase in shear rate when comparing an increase in rectal temperature of 0.5

°C and 1.0 °C (Chiesa *et al.*, 2016). However, shear rate continues to increase by ~100 1/s across 20 minutes when rectal temperature remains constant at 38.5 °C during prolonged hot water immersion (Francisco *et al.*, 2021). A longer duration to reach a higher rectal temperature may contribute to the differences observed in the study of Chiesa *et al.* (2016) compared to the present study where a 0.5 °C greater increase in rectal temperature in the 40-Shoulder condition did not result in difference in superficial femoral shear rate when compared to the 40-Waist condition ( $\Delta 12$  1/s) that had a similar increase in thigh skin temperature ( $\Delta 0.1$  °C). Indeed, a significant strength of the present work is the two duration-matched conditions with similar increases in rectal temperature (i.e. 40-Shoulder and 42-Waist), which allow for the effects of rectal temperature to be examined without being confounded by differences in heating duration.

In a non-submerged limb (e.g. the arms), elevations in rectal temperature during waist-deep immersion, had a systemic effect resulting in increased brachial artery shear rate by 206 1/s and 314 1/s in the 40-Waist and 42-Waist conditions, respectively. These findings are in accordance with previous work, albeit with a longer immersions duration of 45 minutes, demonstrating that knee and ankle-deep immersion increases brachial artery shear rate by ~300 – 600 1/s, when accompanied by ~ 0.5 °C elevations in core body temperature (Cheng *et al.*, 2021). The addition of muscle temperature measurements in the arms and legs would have provided a more comprehensive characterisation of peripheral temperature and enabled greater discussion of the effects of local versus systemic temperature elevation, however these data clearly demonstrate a systemic effect of lower body heating.

#### *Inflammatory responses*

Plasma IL-6 concentrations did not differ following 30-minutes of hot water immersion, whilst cortisol concentrations reduced. Both IL-6 and cortisol concentrations have previously been shown to acutely increase following passive heating protocols (Leicht *et al.*, 2015; Hoekstra *et al.*, 2018, 2021; Mansfield *et al.*, 2021; Steward *et al.*, 2024). However, the duration of heating in these studies is at least 60 minutes compared to 30 minutes in the present study. Cortisol appears to have a biphasic response to heating, whereby there is an initial decrease in concentration followed by a subsequent increase with heating protocols longer than 30 minutes (Ježová *et al.*, 1994; Leicht *et al.*, 2015; Steward *et al.*, 2024). With exercise, there is an exponential relationship between duration and changes in IL-6 (Fischer, 2006) and, although the cause of increases in IL-6 concentration may differ between exercise and passive heating, a similar relationship may exist with passive heating. In support of this, observations from our

laboratory have shown relatively small ( $\sim 0.3$  pg/mL) increases in IL-6 concentrations after 60 but not 30 minutes of arms in 40 °C water immersion (Steward *et al.*, 2024). Whilst IL-6 did not show a significant effect of time in the present study, the partial eta squared was 0.17, with a p value of 0.06, suggesting that the present data was potentially underpowered to show this effect. Regardless of this, the  $0.11 \pm 0.44$  pg/mL increase in IL-6 concentration observed in the present study is of smaller magnitude than previously observed with similar duration moderate-intensity exercise ( $\Delta \sim 0.2$  pg/mL) (Cullen *et al.*, 2016), or greater heating durations ( $\Delta \sim 0.4 - 1.0$  pg/mL) (Mansfield *et al.*, 2021; Hoekstra *et al.*, 2021; Steward *et al.*, 2024). Importantly, the present study showed that alterations to the water temperature or depth of immersion did not alter the acute inflammatory response. Therefore, when considered in the context of the existing literature, duration appears to be a key factor in stimulating an acute inflammatory response to hot water immersion. Indeed, whilst repeated 30-minute heating bouts can elicit vascular or thermoregulatory adaptation (Carter *et al.*, 2014b; Bailey *et al.*, 2016), longer heating durations may be required to induce the IL-6 signalling pathway, such as activation of anti-inflammatory cytokines (e.g. interleukin-10) and assist with the prevention of chronic inflammatory and metabolic diseases. However, longer exposure durations may come at the expense of enjoyment and tolerability.

#### *Tolerability, enjoyment and adverse responses*

The present study showed 30-minutes of immersion was tolerated by all participants in all conditions. However, conditions with larger increases in rectal temperature (40-Shoulder & 42-Waist), through increasing either immersion depth or water temperature, demonstrated poorer perceptual responses and a greater desire to exit. Moreover, 45% of participants reported mild – moderate symptoms of dizziness upon standing following immersion in these conditions. This reinforces the need for caution in naïve users of hot water immersion and the need for more research to identify predictors of standing dizziness. The 40-Waist condition in the present study demonstrated the least negative perceptual responses. Whilst this may lead to longer self-selected durations, or better adherence to a long-term intervention as suggested by Hoekstra *et al.* (2021), our data show that these improved perceptual responses come at the cost of attenuated shear stress and thermal stimuli required for chronic adaptation. However, an unexpected observation was that thermal comfort improved rapidly at the end of immersion in the 40-Shoulder condition when participants had moved onto the stool and removed their arms from the water. This may provide support for the implementation of an immersion protocol where immersion depth begins with maximal body coverage but is reduced after an initial

period (e.g. as performed in Brunt *et al.* (2016)) to extend the immersion duration without it becoming intolerable, thereby attempting to maximise the trade-off between beneficial physiological and favourable perceptual responses.

### *Perspectives*

The observed vascular effects in the present study have important practical implications for the design and application of hot water immersion protocols. This study showed that increases in shear rate are more responsive to changes in local temperature than core body temperature, meaning that limb submersion results in the largest effect on arterial haemodynamic responses, whilst increasing water temperature augments this response in both submerged and non-submerged limbs. These vessel and limb-specific responses, mean that protocols described in insufficient detail (e.g. depth of immersion, immersion of limbs) may be at risk of inaccurate generalisations across vascular beds exposed to different local temperatures and erroneous interpretations of acute haemodynamic effects of a given protocol.

The acute responses shown in the present study are essential for long term adaptation with repeated exposures (Carter *et al.*, 2014b; Dawson *et al.*, 2018; Roxburgh *et al.*, 2023) and suggest that for systemic adaptation, maximising immersion depth or surface area of the body heated is important for heating protocol selection, whilst local vascular adaptation may be better served through a hotter water temperature with a reduced immersion depth. The magnitude of these differences for shear rate was 81 1/s in the brachial artery and 59 1/s in the superficial femoral artery between the 40-Shoulder and 42-Waist conditions. Based on the correlations observed by Dawson *et al.* (2018), this would equate to a ~2-4% greater increase in flow-mediated dilation if these stimuli were repeated as a measure of improved endothelial function and vascular health. Moreover, the present findings on limb/vascular bed specific responses may be relevant for individuals seeking vascular adaptation who may implement hot water immersion differently depending on the requirements of their health condition. For example, patients with claudication of the lower limbs, as seen in people with peripheral artery disease, may favour waist-deep immersion in a hotter water temperature for increases in lower limb shear stress and blood flow as previously investigated by Thomas *et al.* (2017). In contrast, in the general population brachial artery endothelial function is reflective of systemic vascular health (Broxterman *et al.*, 2019) and therefore a greater immersion depth may be more beneficial. This is also supported by the present blood pressure data with the greatest acute reduction in the 40-Shoulder condition of – 10 mmHg for DBP, which would equate to a

chronic reduction of  $\sim$ -2 - -5 mmHg with repeated exposure based on the data observed by Roxburgh *et al.* (2023). Accordingly, these data may be useful to inform future interventional research and practical implementation, however, the present data were collected from young, healthy volunteers and more research is required to investigate the potential different vascular adaptive effects of these immersion protocols in longitudinal interventions with different population groups.

#### Summary

This study provides a strong basis for elucidating different physiological responses to alterations in hot water immersion protocols and demonstrates the potential trade-off between the thermal and arterial haemodynamic stimuli and perceptual responses. Increases in water temperature and immersion depth during 30-minute immersion protocols do not result in different effects on inflammation but have distinct region-specific arterial haemodynamic effects. Beneficial physiological responses may be accompanied with less favourable perceptual responses demonstrating a potential trade-off that should be considered during both acute and chronic protocol implementation.

#### References

- American College of Sports Medicine (2020). *ACSM's Guidelines for Exercise Testing and Prescription*. Lippincott Williams & Wilkins.
- Amin SB, Hansen AB, Mugele H, Willmer F, Gross F, Reimeir B, Cornwell WK, Simpson LL, Moore JP, Romero SA & Lawley JS (2021). Whole body passive heating versus dynamic lower body exercise: a comparison of peripheral hemodynamic profiles. *Journal of Applied Physiology* **130**, 160–171.
- Bailey T, Cable N, Miller G, Sprung V, Low D & Jones H (2016). Repeated Warm Water Immersion Induces Similar Cerebrovascular Adaptations to 8 Weeks of Moderate-Intensity Exercise Training in Females. *Int J Sports Med* **37**, 757–765.
- Broxterman RM et al. (2019). Strong Relationship Between Vascular Function in the Coronary and Brachial Arteries. *Hypertension* **74**, 208–215.
- Brunt VE, Howard MJ, Francisco MA, Ely BR & Minson CT (2016). Passive heat therapy improves endothelial function, arterial stiffness and blood pressure in sedentary humans. *The Journal of Physiology* **594**, 5329–5342.
- Carter HH, Spence AL, Atkinson CL, Pugh CJA, Cable NT, Thijssen DHJ, Naylor LH & Green DJ (2014a). Distinct Effects of Blood Flow and Temperature on Cutaneous Microvascular Adaptation: *Medicine & Science in Sports & Exercise* **46**, 2113–2121.

567 Carter HH, Spence AL, Atkinson CL, Pugh CJA, Naylor LH & Green DJ (2014b). Repeated  
568 core temperature elevation induces conduit artery adaptation in humans. *Eur J Appl*  
569 *Physiol* **114**, 859–865.

570 Charkoudian N, Hart ECJ, Barnes JN & Joyner MJ (2017). Autonomic control of body  
571 temperature and blood pressure: influences of female sex hormones. *Clin Auton Res* **27**,  
572 149–155.

573 Chaseling GK, Debray A, Gravel H, Ravanelli N, Bartlett A-A & Gagnon D (2023). The acute  
574 effect of heat exposure on forearm macro- and microvascular function: Impact of  
575 measurement timing, heating modality and biological sex. *Experimental Physiology*  
576 **108**, 221–239.

577 Cheng JL, Williams JS, Hoekstra SP & MacDonald MJ (2021). Improvements in vascular  
578 function in response to acute lower limb heating in young healthy males and females.  
579 *Journal of Applied Physiology* **131**, 277–289.

580 Chiesa ST, Trangmar SJ & González-Alonso J (2016). Temperature and blood flow  
581 distribution in the human leg during passive heat stress. *Journal of Applied Physiology*  
582 **120**, 1047–1058.

583 Convertino VA (2014). Neurohumoral mechanisms associated with orthostasis: reaffirmation  
584 of the significant contribution of the heart rate response. *Front Physiol*; DOI:  
585 10.3389/fphys.2014.00236.

586 Coris EE, Walz SM, Duncanson R, Ramirez AM & Roetzheim RG (2006). Heat illness  
587 symptom index (HISI): a novel instrument for the assessment of heat illness in athletes.  
588 *Southern Medical Journal* **99**, 340–346.

589 Cullen T, Clarke ND, Hill M, Menzies C, Pugh CJA, Steward CJ & Thake CD (2020). The  
590 health benefits of passive heating and aerobic exercise: To what extent do the  
591 mechanisms overlap? *Journal of Applied Physiology* **129**, 1304–1309.

592 Cullen T, Thomas AW, Webb R & Hughes MG (2016). Interleukin-6 and associated cytokine  
593 responses to an acute bout of high-intensity interval exercise: the effect of exercise  
594 intensity and volume. *Appl Physiol Nutr Metab* **41**, 803–808.

595 Dawson EA, Cable NT, Green DJ & Thijssen DHJ (2018). Do acute effects of exercise on  
596 vascular function predict adaptation to training? *Eur J Appl Physiol* **118**, 523–530.

597 Dill DB & Costill DL (1974). Calculation of percentage changes in volumes of blood, plasma,  
598 and red cells in dehydration. *Journal of Applied Physiology* **37**, 247–248.

599 Ely BR, Clayton ZS, McCurdy CE, Pfeiffer J, Needham KW, Comrada LN & Minson CT  
600 (2019). Heat therapy improves glucose tolerance and adipose tissue insulin signaling in  
601 polycystic ovary syndrome. *American Journal of Physiology-Endocrinology and*  
602 *Metabolism* **317**, E172–E182.

603 Epstein Y & Moran DS (2006). Thermal Comfort and the Heat Stress Indices. *Ind Health* **44**,  
604 388–398.



605 Fischer CP (2006). Interleukin-6 in acute exercise and training: what is the biological  
606 relevance? *Exerc Immunol Rev* **12**, 6–33.

607 Francisco MA, Colbert C, Larson EA, Sieck DC, Halliwill JR & Minson CT (2021).  
608 Hemodynamics of post-exercise vs. post hot water immersion recovery. *J Appl Physiol*  
609 (1985); DOI: 10.1152/jappphysiol.00260.2020.

610 Gibson OR, Laitano O, Watanabe K & González-Alonso J (2025). Differential intestinal injury  
611 and unchanged systemic inflammatory responses to leg and whole-body passive  
612 hyperthermia in healthy humans. *Exp Physiol*; DOI: 10.1113/EP092389.

613 Hafen PS, Preece CN, Sorensen JR, Hancock CR & Hyldahl RD (2018). Repeated exposure to  
614 heat stress induces mitochondrial adaptation in human skeletal muscle. *Journal of*  
615 *Applied Physiology* **125**, 1447–1455.

616 Hashimoto M, Akishita M, Eto M, Ishikawa M, Kozaki K, Toba K, Sagara Y, Taketani Y,  
617 Orimo H & Ouchi Y (1995). Modulation of Endothelium-Dependent Flow-Mediated  
618 Dilatation of the Brachial Artery by Sex and Menstrual Cycle. *Circulation* **92**, 3431–  
619 3435.

620 Heinonen I, Brothers RM, Kemppainen J, Knuuti J, Kalliokoski KK & Crandall CG (2011).  
621 Local heating, but not indirect whole body heating, increases human skeletal muscle  
622 blood flow. *Journal of Applied Physiology* **111**, 818–824.

623 Hesketh K, Shepherd SO, Strauss JA, Low DA, Cooper RJ, Wagenmakers AJM & Cocks M  
624 (2019). Passive heat therapy in sedentary humans increases skeletal muscle  
625 capillarization and eNOS content but not mitochondrial density or GLUT4 content.  
626 *American Journal of Physiology-Heart and Circulatory Physiology* **317**, H114–H123.

627 Hoekstra SP, Bishop NC, Faulkner SH, Bailey SJ & Leicht CA (2018). Acute and chronic  
628 effects of hot water immersion on inflammation and metabolism in sedentary,  
629 overweight adults. *Journal of Applied Physiology* **125**, 2008–2018.

630 Hoekstra SP, Bishop NC & Leicht CA (2020). Elevating body temperature to reduce chronic  
631 low-grade inflammation: a welcome strategy for those unable to exercise? **26**, 42–55.

632 Hoekstra SP, Ogawa T, Dos Santos M, Handsley G, Bailey SJ, Goosey-Tolfrey VL, Tajima F,  
633 Cheng JL & Leicht CA (2021). The effects of local versus systemic passive heating on  
634 the acute inflammatory, vascular and glycaemic response. *Appl Physiol Nutr Metab* **46**,  
635 808–818.

636 Ježová D, Kvetňanský R & Vigaš M (1994). Sex differences in endocrine response to  
637 hyperthermia in sauna. *Acta Physiologica Scandinavica* **150**, 293–298.

638 Kim K, Monroe JC, Gavin TP & Roseguini BT (2020). Skeletal muscle adaptations to heat  
639 therapy. *Journal of Applied Physiology*; DOI: 10.1152/jappphysiol.00061.2020.

640 Knief U & Forstmeier W (2020). Violating the normality assumption may be the lesser of two  
641 evils. 498931. Available at: <https://www.biorxiv.org/content/10.1101/498931v2>  
642 [Accessed May 27, 2022].

643 Koch Esteves N, McDonald J & González-Alonso J (2024). Thermo-haemodynamic coupling  
644 during regional thigh heating: Insight into the importance of local thermosensitive  
645 mechanisms in blood circulation. *Experimental Physiology* **109**, 600–613.

646 Leicht CA, Kouda K, Umemoto Y, Banno M, Kinoshita T, Moriki T, Nakamura T, Bishop NC,  
647 Goosey-Tolfrey VL & Tajima F (2015). Hot water immersion induces an acute cytokine  
648 response in cervical spinal cord injury. *Eur J Appl Physiol* **115**, 2243–2252.

649 Liu C-T & Brooks GA (2011). Mild heat stress induces mitochondrial biogenesis in C2C12  
650 myotubes. *Journal of Applied Physiology* **112**, 354–361.

651 Low DA, Keller DM, Wingo JE, Brothers RM & Crandall CG (2011). Sympathetic nerve  
652 activity and whole body heat stress in humans. *Journal of Applied Physiology* **111**,  
653 1329–1334.

654 Mansfield RG, Hoekstra SP, Bill JJ & Leicht CA (2021). Local cooling during hot water  
655 immersion improves perceptions without inhibiting the acute interleukin-6 response.  
656 *Eur J Appl Physiol*; DOI: 10.1007/s00421-021-04616-5.

657 Monroe JC, Song Q, Emery MS, Hirai DM, Motaganahalli RL & Roseguini BT (2021). Acute  
658 effects of leg heat therapy on walking performance and cardiovascular and  
659 inflammatory responses to exercise in patients with peripheral artery disease.  
660 *Physiological Reports* **8**, e14650.

661 Nash D, Hughes MG, Butcher L, Aicheler R, Smith P, Cullen T & Webb R (2023). IL-6  
662 signaling in acute exercise and chronic training: Potential consequences for health and  
663 athletic performance. *Scandinavian Journal of Medicine & Science in Sports* **33**, 4–19.

664 Obi S, Nakajima T, Hasegawa T, Kikuchi H, Oguri G, Takahashi M, Nakamura F, Yamasoba  
665 T, Sakuma M, Toyoda S, Tei C & Inoue T (2017). Heat induces interleukin-6 in skeletal  
666 muscle cells via TRPV1/PKC/CREB pathways. *Journal of Applied Physiology* **122**,  
667 683–694.

668 Oi H, Tabata K, Naka Y, Takeda A & Tochihara Y (2012). Effects of heated seats in vehicles  
669 on thermal comfort during the initial warm-up period. *Applied Ergonomics* **43**, 360–  
670 367.

671 Rhind SG, Gannon GA, Shephard RJ, Buguet A, Shek PN & Radomski MW (2004). Cytokine  
672 induction during exertional hyperthermia is abolished by core temperature clamping:  
673 neuroendocrine regulatory mechanisms. *International Journal of Hyperthermia* **20**,  
674 503–516.

675 Roxburgh BH, Campbell HA, Cotter JD, Reymann U, Williams MJA, Gwynne-Jones D &  
676 Thomas KN (2023). Acute and adaptive cardiovascular and metabolic effects of passive  
677 heat therapy or high-intensity interval training in patients with severe lower-limb  
678 osteoarthritis. *Physiological Reports* **11**, e15699.

679 Shin YO, Lee JB, Min YK & Yang HM (2013). Heat acclimation affects circulating levels of  
680 prostaglandin E2, COX-2 and orexin in humans. *Neuroscience Letters* **542**, 17–20.

681 Steward CJ, Hill M, Menzies C, Bailey SJ, Rahman M, Thake CD, Pugh CJA & Cullen T  
682 (2024). Post exercise hot water immersion and hot water immersion in isolation

683 enhance vascular, blood marker, and perceptual responses when compared to exercise  
684 alone. *Scandinavian Journal of Medicine & Science in Sports* **34**, e14600.

685 Steward CJ, Menzies C, Clarke ND, Harwood AE, Hill M, Pugh CJA, Thake CD & Cullen T  
686 (2023). The effect of age and mitigation strategies during hot water immersion on  
687 orthostatic intolerance and thermal stress. *Experimental Physiology* **108**, 554–567.

688 Thomas KN, van Rij AM, Lucas SJE & Cotter JD (2017). Lower-limb hot-water immersion  
689 acutely induces beneficial hemodynamic and cardiovascular responses in peripheral  
690 arterial disease and healthy, elderly controls. *American Journal of Physiology-  
691 Regulatory, Integrative and Comparative Physiology* **312**, R281–R291.

692 Watanabe K, Koch Esteves N, Gibson OR, Akiyama K, Watanabe S & González-Alonso J  
693 (2024). Heat-related changes in the velocity and kinetic energy of flowing blood  
694 influence the human heart's output during hyperthermia. *J Physiol* **602**, 2227–2251.

695 Welc SS, Phillips NA, Oca-Cossio J, Wallet SM, Chen DL & Clanton TL (2012). Hyperthermia  
696 increases interleukin-6 in mouse skeletal muscle. *Am J Physiol Cell Physiol* **303**, C455–  
697 C466.

698 Woodman RJ, Playford DA, Watts GF, Cheetham C, Reed C, Taylor RR, Puddey IB, Beilin  
699 LJ, Burke V, Mori TA & Green D (2001). Improved analysis of brachial artery  
700 ultrasound using a novel edge-detection software system. *Journal of Applied  
701 Physiology* **91**, 929–937.